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BOX: PATENT APPLICATION

SIR:

Transmitted herewith for filing is the patent application (including Specification, Claims, and Abstract (47 pages) of:

Inventor(s): **Maqsudul Alam and Randy Larsen**For : **HEME PROTEINS HEMAT-HS AND HEMAT-BS AND THEIR USE IN
MEDICINE AND MICROSENSORS*****If a CONTINUING APPLICATION, please mark where appropriate and supply the requisite information below and in a preliminary amendment:*☐ continuation ☐ divisional ☐ Continuation-In-Part (CIP)
of prior application Serial No. _____Prior application information: Examiner :
Art Unit :

Enclosed are:

- ☒ 8 sheets of informal drawings.
- ☐ **Signed** Combined Declaration and Power of Attorney (____ pages).
- ☐ **Copy of signed** Combined Declaration and Power of Attorney (____ pages) from a prior application (1.63(d) (for continuation/divisional).
- ☐ **Signed** statement deleting inventor(s) named in prior application (____ pages) (1.63(d)(2) and 1.33(b)).
- ☐ **Incorporation By Reference:** The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied herewith, is considered as being part of the disclosure of the enclosed application and is hereby incorporated by reference therein.
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- ☐ Assignment Transmittal Letter.
- ☐ Certified copy of a foreign priority document.
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- ☐ Verified statement to establish small entity status (____ pages) (newly signed or copy filed in prior application).



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- ☐ Preliminary Amendment (____ pages).
- ☐ Information Disclosure Statement, form PTO-1449 (____ pages) and ____ references.
- ☒ **UNSIGNED** Combined Declaration and Power of Attorney (2 pages).
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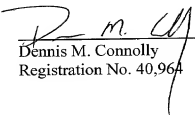
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DOCKET NO. : **201040/1020**

APPLICANT(S) : **Maqsudul Alam and Randy Larsen**

TITLE : **HEME PROTEINS HEMAT-*HS* AND HEMAT-*BS* AND THEIR
USE IN MEDICINE AND MICROSENSORS**

Certificate is attached to the **Utility Patent Application Transmittal Form
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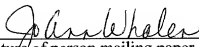
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(Signature of person mailing paper or fee)

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**TITLE: HEME PROTEINS HemAT-*Hs*
AND HemAT-*Bs* AND THEIR USE
IN MEDICINE AND
MICROSENSORS**

**INVENTORS: MAQSUDUL ALAM AND RANDY
LARSEN**

DOCKET: 201040/1020

HEME PROTEINS HEMAT-*HS* AND HEMAT-*BS* AND THEIR USE IN MEDICINE AND MICROSENSORS

The subject matter of this application was made with support from the United States Government under Grant No. MSB960086 from the National Science Foundation. The United States Government may retain certain rights.

BACKGROUND OF THE INVENTION

Heme proteins such as hemoglobin and myoglobin play an essential role in stabilizing molecular oxygen for transport and storage. The oxygen carrying portion of the red blood cell is hemoglobin, a tetrameric protein molecule composed of two identical alpha globins (alpha 1, alpha 2), two identical beta globins (beta 1, beta 2) and four heme molecules. A heme molecule is incorporated into each of the alpha and beta globins to give alpha and beta subunits. Heme is a macrocyclic organic molecule that contains an iron atom at its center; each heme can combine reversibly with one ligand molecule, for example oxygen. In a hemoglobin tetramer, each alpha subunit is associated with a beta subunit to form two stable alpha/beta dimers, which in turn associate to form the tetramer (a homodimer). The subunits are noncovalently associated through Van der Waals forces, hydrogen bonds and salt bridges. Ligands, particularly oxygen, bind reversibly to the reduced form of the iron (ferrous, Fe^{2+}) in the heme. Other ligands which compete with oxygen for the heme group include carbon monoxide and nitric oxide.

It is not always practical to transfuse a patient with donated blood. The well known complications of blood transfusion namely incompatibility reactions, disease transmission, immunosuppression and the storage limitations of erythrocytes points to the need for the development of blood substitutes devoid of these shortcomings. In these situations, use of a red blood cell substitute is necessary. A "blood substitute" is a preparation that does not necessarily replace blood in all of its functions, but an emergency resuscitative fluid that is capable of efficiently transporting oxygen to tissue. This fluid, however, must be free of toxic side-effects, as well as of agents of disease such as bacteria and viruses.

For over 50 years, efforts directed to the development of a blood substitute have focused on hemoglobin (Hb). Hemoglobin (Hgb) is the oxygen-carrying component of

blood. Hemoglobin circulates through the bloodstream inside small enucleate cells called erythrocytes (red blood cells). Hemoglobin is a protein constructed from four associated polypeptide chains, and bearing prosthetic groups known as hemes. The erythrocyte helps maintain hemoglobin in its reduced, functional form. The heme iron atom is labile to oxidation, but may be reduced again by one of two enzyme systems within the erythrocyte, the cytochrome b5 and glutathione reduction systems.

Hemoglobin exhibits cooperative binding of oxygen by the four subunits of the hemoglobin molecule (two alpha-globins and two beta-globins in the case of HbA), and this cooperativity greatly facilitates efficient oxygen transport. Cooperativity, achieved by the so-called heme-heme interaction, allows hemoglobin to vary its affinity for oxygen. Hemoglobin reversibly binds up to four moles of oxygen per mole of Hb. At high oxygen concentration, such as that found in the lungs, the oxygen affinity is high and hemoglobin is almost saturated with oxygen. At low oxygen concentration, such as that found in actively respiring tissue, the oxygen affinity is lowered and oxygen is unloaded. The oxygen affinity of hemoglobin is lowered by the presence of 2,3-diphosphoglycerate (2,3-DPG), chloride ions and hydrogen ions. Respiring tissue releases carbon dioxide into the blood and lowers its pH (i.e. increases the hydrogen ion concentration), thereby causing oxygen to dissociate from hemoglobin and allowing it to diffuse into individual cells.

The ability of hemoglobin to alter its oxygen affinity, increasing the efficiency of oxygen transport around the body, is dependent on the presence of the metabolite 2,3-DPG. Inside the erythrocyte 2,3-DPG is present at a concentration nearly as great as that of hemoglobin itself. In the absence of 2,3-DPG "conventional" hemoglobin binds oxygen very tightly and would release little oxygen to respiring tissue.

Aging erythrocytes release small amounts of free hemoglobin into the blood plasma where it is rapidly bound by the scavenging protein haptoglobin. The hemoglobin-haptoglobin complex is removed from the blood and degraded by the spleen and liver.

It is clear from the above considerations that free native hemoglobin A, injected directly into the bloodstream, would not support efficient oxygen transport about the body. The essential allosteric regulator 2,3-DPG is not present in sufficient concentration in the plasma to allow hemoglobin to release much oxygen at venous oxygen tension, and

free hemoglobin would be rapidly inactivated as an oxygen carrier by auto-oxidation of the heme iron.

Therefore, a need exists for a substitute other than hemoglobin which can bind and carry oxygen to cells. This substitute may also be used in other applications where hemoglobin is used, including as a biological sensor for oxygen. The present invention provides proteins which meet that need.

SUMMARY OF THE INVENTION

The present invention provides isolated archaeal and bacterial heme binding proteins which reversibly bind oxygen with a low affinity.

The invention also provides a blood substitute containing the bacterial heme binding protein which reversibly binds oxygen with a low affinity.

Another embodiment of the invention is a method for controlled storage of oxygen. A bacterial heme binding protein which reversibly binds oxygen with a low affinity is contacted with oxygen allowing the protein to bind and store oxygen. The invention also provides a method of sensing gaseous ligands. A heme binding bacterial protein is exposed to a test sample and a change in the conformation of the protein is measured.

Yet another embodiment of the invention is a chimeric protein having a heme-binding domain and a heterologous signaling domain.

The invention further provides an isolated nucleic acid molecule which encodes a heme binding bacterial protein that reversibly binds oxygen with a low affinity.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the conserved sequences within HemAT-*Hs*, HemAT-*Bs*, and sperm-whale myoglobin (SWMb). Black boxes indicate positions at which the residues are identical, and gray boxes highlight residues that are similar. Sequences were aligned using the Clustal program of the MegAlign/DNASTAR package. A) Alignment of the amino-terminal domain of HemAT-*Hs*, HemAT-*Bs*, and SWMb. Helical regions in SWMb (helices A-H) (B. C. Clothia, et al., J. Mol. Biol. 196:199 (1987); S. N. Vinogradov et al., Comp. Biochem. Physiol. 106B:1 (1993), which are hereby

incorporated by reference) are delineated by dotted arrows. Pro (P), Phe (F), and His (H) residues in SWMb that are highly conserved among all globins are marked with asterisks.

B) Alignment of the carboxyl-terminal domains of HemAT-*Hs*, HemAT-*Bs*, and Tsr (B. K. Kendall, et al., Nature 301:623 (1983); G. L. Hazelbauer, Curr. Opin. Struct. Biol., 2:505 (1992), which are hereby incorporated by reference).

Figure 2 is a characterization of HemAT proteins. Figure 2A shows the purified HemAT-*Hs* and HemAT-*Bs* in 10% SDS-PAGE. Approximately 5 µg of purified protein were loaded in each lane for separation during SDS-PAGE in 10% acrylamide (M. Alam et al., J. Bacteriol., 173:5837 (1991), which is hereby incorporated by reference). Lane 1, HemAT-*Hs*; lane 2, HemAT_{6xHis}-*Hs*; and lane 3, HemAT-*Bs*. The MW markers (kDa) are shown at the left. Figure 2B is a fluorograph and immunoblot of HemAT-*Hs*.

Radiolabeling and immunoblotting were performed as previously described (M. Alam et al., J. Bacteriol., 173:5837 (1991), which is hereby incorporated by reference). Lane 1, fluorograph of proteins from the Δ *hemAT-Hs*; lane 2, fluorograph of proteins from the Δ *hemAT-Hs/hemAT-Hs*⁺⁺ strain (A. Brooun, Ph.D thesis. University of Hawaii, Hawaii (1997), which is hereby incorporated by reference). *NdeI* and *XbaI* restriction sites were used to clone the *hemAT-Hs* gene into the shuttle vector pKJ427. Primers introducing flanking *NdeI* and *XbaI* restriction sites were used for PCR amplification. The PCR product was initially cloned into the pCRR-Blunt II TOPO cloning vector and later subcloned into plasmid pKJ427 after digestion with *NdeI* and *XbaI*. The resulting plasmid was introduced into the Δ *hemAT-Hs* strain. Lane 1: immunoblot of Δ *hemAT-Hs* strain; and lane 2: immunoblot of Δ *hemAT-Hs/hemAT-Hs*⁺⁺ strain using anti-transducer peptide antibody (W. Zhang et al., Proc. Natl. Acad. Sci. U.S.A. 93:4649 (1996), which is hereby incorporated by reference). Bars indicate the positions of molecular weight markers (kDa). Radiolabeling and immunoblot experiments were performed according to Alam & Hazelbauer (Alam et al., J. Bacteriol., 173:5837-5842 (1991)).

Figure 3 provides a comparison of the proteins used in the homology analyses. M1 and M2 are the site of myoglobin recognition. M2 is the site of HemAT recognition. The H-box is the primary site of microbial hemoglobin recognition.

Figure 4 shows absorption spectra of purified HemAT-*Hs*, HemAT-*Bs*, and horse-heart myoglobin (HHMb). Panel A shows oxygenated forms of purified HemAT-*Hs*, HemAT-*Bs*, and oxymyoglobin. Panel B shows deoxygenated forms of HemAT-*Hs*,

HemAT-*Bs*, and myoglobin. Panel C shows CO-bound forms of HemAT-*Hs*, HemAT-*Bs*, and myoglobin. Panel D shows reoxidized forms of HemAT-*Hs*, HemAT-*Bs*. Samples concentrations are approximately 20 μ M in heme. Deoxygenated samples were prepared by the addition of sodium dithionite to the deaerated protein solutions.

- 5 Figure 5 shows aerotactic responses in *H. salinarum* and *B. subtilis*. Panel A shows *H. salinarum* strain Flx15 (HtrVIII and HemAT-*Hs* present), and mutant strains Δ hemAT-*Hs* (HtrVIII present) and Δ htrVIII (HemAT-*Hs* present). Panel B shows wild-type *B. subtilis* strain OI1085 and mutant strains OI3545 (Δ ten) and OI3555 (overexpression of hemAT-*Bs* in Δ ten). All cells were grown to mid-logarithmic phase.
- 10 Microcapillaries (internal dimension 100x10 μ m) were filled halfway with cell suspension. The capillaries were sealed at both ends and placed on a microscope stage prewarmed to 35-37 $^{\circ}$ C. Time-lapse, dark-field microscopic images were recorded using a video-digitized camera linked to a computer. The images shown were taken at 180 min for *H. salinarum* and at 30 min for *B. subtilis*. The meniscus at the air interface is visible
- 15 to the right in each image.

Figure 6 provides transient absorption data subsequent to CO photolysis obtained at 430 nm at 25 $^{\circ}$ C and 1 atm CO for HemAT-*Hs* (solid line) and HemAT-*Bs* (dotted line). Samples were approximately 20 μ M. The traces are the average of 50 laser pulses (532 nm excitation, 7 ns pulse width, 10 mJ/pulse).

- 20 Figure 7 shows the transient difference spectrum (25 μ s subsequent to photolysis) overlaid with the equilibrium difference spectrum (deoxy minus CO-bound) for HemAT-*Hs* (top panel) and HemAT-*Bs* (bottom panel). Sample conditions are as described in Figure 4.

- 25 Figure 8 provides CO-off rate data for HemAT-*Hs* (dashed line), HemAT-*Bs* (dotted line), and horse heart Mb (solid line). Changes in absorbance as a function of time at 418 were monitored after the addition of potassium ferricyanide (final concentration of 1.5 mM). See Example 17 for details.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides an isolated bacterial myoglobin-like heme binding protein which reversibly binds oxygen with a low affinity. These proteins are a new class of heme proteins that bind diatomic oxygen through their prosthetic group and trigger negative aerotactic responses. HemAT-*Hs* and HemAT-*Bs* are the first myoglobin-like heme proteins in the *Archaea* and in *Bacteria*, respectively. Purified HemAT-*Hs* and HemAT-*Bs* exhibit spectral properties similar to oxygen-bound myoglobin.

Deoxygenation of either protein results in absorption shifts similar to those observed for deoxymyoglobin. The oxy-/deoxy spectral changes in HemAT-*Hs* and in HemAT-*Bs* are completely reversible, a characteristic feature of the heme prosthetic group in myoglobin. The C-terminus of both proteins has high homology with the signaling domain of bacterial methyl-accepting chemoreceptors and they mediate aerotaxis. By site-directed mutagenesis the fifth coordination site of the heme iron was identified in HemAT-*Hs* and HemAT-*Bs* comparable to myoglobin.

In a preferred embodiment of the invention, the isolated heme-binding protein has both a heme binding domain and a signaling domain.

When the *hemAT-Hs* gene was originally cloned, its product was predicted to be a soluble signal transducer (W. Zhang et al., Proc. Natl. Acad. Sci. U.S.A. 93:4649 (1996), which is hereby incorporated by reference). HemAT-*Bs* was identified in the *B. subtilis* genome-sequencing project as the product of an open-reading frame encoding a protein with marked similarities to methyl-accepting chemotaxis proteins (MCP) (F. Kunst et al., Nature 390:249 (1997), which is hereby incorporated by reference). The predicted translation products of the *hemAT-Hs* and *hemAT-Bs* genes, comprising 489 and 432 residues, respectively, exhibit two striking features: a) their amino-termini (residues 1-184 in HemAT-*Hs* and 1-175 in HemAT-*Bs*) display limited homology to myoglobin (Figure 1A); b) residues 222 to 489 of HemAT-*Hs* and 198 to 432 of HemAT-*Bs* are 30% identical to the cytoplasmic signaling domain of Tsr, an MCP from *Escherichia coli* (Figure 1B).

The residues absolutely conserved among all globins are the proximal His in the F helix (F8) and Phe in the CD region (CD1) (B. C. Clothia, et al., J. Mol. Biol. 196:199 (1987); S. N. Vinogradov et al., Comp. Biochem. Physiol. 106B:1 (1993), which are hereby incorporated by reference). Highly conserved residues include the distal His in

the E helix (E7), Phe in the CD4 region, and Pro at the beginning of the C helix (C2) (B. C. Clothia, et al., J. Mol. Biol., 196:199 (1987); S. N. Vinogradov et al., Comp. Biochem. Physiol., 106B:1 (1993), which are hereby incorporated by reference). Three of these residues (Pro in C2, Phe in CD1, His in F8) are conserved in both HemAT-*Hs* and HemAT-*Bs* (asterisks in Figure 1A). These features suggested to us that HemATs may be heme-containing proteins that generate signals in response to binding of oxygen.

Both proteins, HemAT-*Hs* and HemAT-*Bs*, can be expressed in *E. coli* from recombinant vectors. PCR primers with sequences flanking the *hemAT-Hs* gene from *H. salinarum* strain Flx15 and encoding a *NdeI* or *BamHI* restriction site were used to amplify and clone the chromosomal gene into the pET expression vector (Novogen Inc.). The PCR product was initially ligated into the pCR^R-Blunt II TOPO cloning vector (Invitrogen, Inc.) and then subcloned into pET-3b after digestion of the donor and recipient plasmids with *NdeI* and *BamHI*. The resulting plasmid was introduced into the *E. coli* pLysS strain for protein expression. PCR primers with sequences flanking the *hemAT-Bs* gene from *B. subtilis* strain OI1085 and encoding a *BamHI* or *PstI* restriction site were used to amplify and clone the chromosomal gene into the pCR^R-Blunt II TOPO vector. This fragment was later subcloned into the pMALcII expression vector (New England Biolabs, Inc.), which was introduced into the *E. coli* pLysS strain for protein expression.

Recombinant HemAT-*Hs* is purified using anion-exchange and gel-filtration chromatography. BL21 pLysS host cells harboring plasmids carrying the *hemAT-Hs* or *hemAT-Bs* genes were grown in Luria-Bertani broth with appropriate antibiotics, and synthesis of the proteins was induced with 0.6 mM isopropyl-D-thiogalactopyranoside. After a two hour induction, the cells were harvested by low speed centrifugation (4000 x g) at 4°C for 15 min. The pellets were resuspended in buffer (50 mM NaCl, 50 mM Tris-HCl, pH 6.0) and sonicated for 4 min (12 pulses of 20 sec with 30 sec pauses). The cell lysate was centrifuged at 28,000 x g for 20 min. The red supernatant became the source of proteins for purification. The HemAT-*Hs* supernatant was applied to an anion-exchange POROS HQ/M column equilibrated with buffer (50 mM Tris-HCl, pH 6.0). A linear gradient of NaCl (0-1500 mM) was applied, and HemAT-*Hs* eluted at about 400 mM. Fractions containing HemAT-*Hs* (monitored by the Soret band absorbance at 410 nm and SDS-PAGE) were concentrated and applied to a HiLoad Superdex 200 gel-

filtration column. Fractions containing HemAT-*Hs* were concentrated with an Amicon 100K concentrator.

A saturated $(\text{NH}_4)_2\text{SO}_4$ solution was added to 30% saturation to the HemAT-*Bs* supernatant and centrifuged at $28,000 \times g$ for 20 min. The optically clear, light-red supernatant was further fractionated by adding $(\text{NH}_4)_2\text{SO}_4$ to 36% saturation, and the precipitate was pelleted by centrifugation. The pellet was resuspended in buffer (200 mM NaCl, 50 mM Tris-HCl, pH 8.0) and applied to a HiLoad Superdex 75 column. Fractions containing HemAT-*Bs* were concentrated with an Amicon 50K concentrator.

During SDS-polyacrylamide gel electrophoresis (SDS-PAGE), purified HemAT-*Hs* migrates slower than expected from its calculated molecular mass of 52.8 kDa (Figure 2A, lane 1). This behavior is consistent with the highly acidic nature of many halophilic proteins (K. Ihara et al., Arch. Biochem. Biophys., 286:111 (1991), which is hereby incorporated by reference). HemAT-*Hs* is purified from *H. salinarum* by metal-affinity and gel-filtration chromatography as a recombinant protein (HemAT_{6xHis}-*Hs*) carrying a carboxyl-terminal six-histidine tag (Figure 2A, lane 2). A plasmid encoding carboxyl-terminal 6 His-tagged HemAT-*Hs* was constructed by two-step PCR. In the first step, 6 His codons were fused to *hemAT-Hs* immediately in front of the natural stop codon. In the second step, a *Xba*I restriction site was introduced at the 3' end of the gene. The second PCR product was subcloned into the *Nde*I and *Xba*I sites of plasmid pKJ427. This plasmid was introduced into a Δ *hemAT-Hs* strain (A. Brooun et al., J. Bacteriol., 180:1642 (1998), which is hereby incorporated by reference). Cells grown at 39 °C to mid-logarithmic phase were harvested by centrifugation ($4000 \times g$) at 4 °C. The pellet was resuspended in buffer (200 mM NaCl, 50 mM Tris-HCl, pH 8.0) and sonicated for 3 min (12 pulses of 15 sec with 20 sec pauses). The cell lysate was centrifuged ($100,000 \times g$) at 14 °C for 30 min, and the supernatant was used for purification. The POROS MC/M affinity column was washed with 1 M NaCl, 50 mM EDTA (pH 8.0), charged with 100 mM CoCl_2 , and finally washed with 3M NaCl. The column was equilibrated with buffer (200 mM NaCl, 50 mM Tris-HCl, pH 8.0) prior to loading the sample. HemAT_{6xHis}-*Hs* was eluted with a linear gradient of imidazole (0-250 mM). The peak fractions were collected, concentrated, and applied to a HiLoad Superdex 200 gel-filtration column. The peak fractions were concentrated with an Amicon 100K concentrator. HemAT-*Bs* is purified using a combination of ammonium-sulfate precipitation/fractionation and gel-

filtration chromatography. As expected, purified HemAT-*Bs* migrates during SDS-PAGE as a 48.7 kDa protein (Figure 2A, lane 3).

The preferred bacterial heme binding proteins are myoglobin-like proteins. In particular, the heme binding proteins show greater than 20% identity to a vertebrate myoglobin protein, such as sperm whale myoglobin. More preferred are proteins which show greater than 30% or 50% identity. The level of identity is calculated using the protein alignment program of BLAST with the default parameters.

In a preferred embodiment, the heme-binding protein is isolated from *Archaea*. The *Archaea* are a group of organisms often found in extreme environments, such as high temperatures, high salt concentrations, and acidic conditions. The conditions are often so extreme that other organisms are unable to survive in that environment. Proteins isolated from the *Archaea* often exhibit higher stability in the presence of high temperatures, high salt concentrations, or low pH. Generally, the proteins isolated from *Archaea* are preferred due to their higher stability.

In particular, the protein is isolated from *Halobacterium salinarum*. *H. salinarum* is a salt tolerant organism. Similarly, the HemAT-*Hs* protein is salt tolerant. The sequence of the gene encoding the HemAT-*Hs* protein is shown in SEQ. ID. No. 1 as follows:

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20  ATGAGCAACG ATAATGACAC TCTCGTGACC GCCGACGTTT GGAACGGGAT CGACGGGCAC 60
    GCACTCGCGC ACCGGATCGG CCTCGACGAG GCGGAGATCG CGTGGCGGCT GTCGTTCAAC 120
    GGGATCGAGC ACGACACGAT GGCCGCGCTC GCCGCCGAAC AGCCGCTGTT CGAAGCCACC 180
    GCGGACGCGC TGGTGACCGA CTTCTACGAC CACTTGGAGT CTTACGAGCG CACACAGGAC 240
    CTCTTCGCGA ACTCCACGAA GACCGTCGAG CAACTCAAAG AGACGCGAGC CGAGTACTTG 300
25  CTGGGCCTCG GCGCGGCGGA GTACGACACC GAGTACGCGC CCCAGCGCGC CCGTATCGGG 360
    AAGATACACG ACGTGCTCGG GCTCGGACCG GACGCTCTATC TGGGCGCGTA CACGCGATAC 420
    TACACGGGGC TGTGTGACGC GCTTGCCGAC GACGTGGTGC CCGACCGCGG CGAGGAGGCG 480
    GCCGCGCGCG TCGACGAACT CGTGGCCCGG TTCTCGCCGA TGTGAAGCT GTTGACCTTC 540
    GATCAGCAGA TCGCAATGGA CACCTACATC GACTCGTACG CCCAGCGCCT CCACGACGAG 600
30  ATCGACAGCC GCCAGGAGTT GGCGAACGCG GTCCGCCACG ACGTGAAGC ACCGCTGTCC 660
    TCGCTGGAGG CGACCTCGCA GGACGTCGCC GAGCGCACGG ACACGATGCG GGCCCGCACC 720
    GACGACCAAG TCGACCGCAT GGTGTGACGT AGCCGTGAGA TATCCAGCGT GTCCGCGAGC 780
    GTCGAGGAGG TCGCCTCGAC GGCCGACGAC GTCCGCCGGA CCAGCGAGGA CGCCGAGGCG 840
    CTGGCCCGAC AGGGCGAGGC GGCCGCCGAC GACGCGCTCG CCACGATGAC AGACATCGAC 900
35  GAGGCGACCG ACGGCGTCC CCGGGCGCTG GAACAGCTCG GCGAGCGCGC CGCCGAGGTC 960
    GAATCAGTGA CCGGCGTGAT CGACGACATC GCCGAGCAGA CGAATCATCT GGCCTGAAC 1020
    GCGTCCATCG AGGCGCGCGG CGCCGGGGAG GCGGGCGAGG GGTTCGCGGT CGTCCCGGAC 1080

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GAGGTCAAGG CCCTCGCCGA GGAGTCCCGC GAGCAGTCCA CGCGCGTCGA GGAGCTCGTC 1140
 GAGCAGATGC AGGCGGAGAC CGAGGAGACG GTCGACCACT TGGACGAGGT CAACCCAGCGC 1200
 ATCGGCGAGG GCGTCGAGCG CGTCGAGGAG GCGATGGAGA CCTCCAGGA GATCACCGAC 1260
 GCGCTCGAGG ACGCCGCAAG CCGGATGCAG GAGGTGTCGA CGGCGACCGA CGAACAGGCG 1320
 5 GTGAGCACC GAGAGGTGCG CGAGATGGTC GACGGTGTCT ACGACCGCGC GGGCCGAGATC 1380
 GCGGCCGCC TCGATGACAT CGCCGACGCG ACCGATCAGC AGGTCCGACG CGTCGAGGAG 1440
 GTCGCGGAGA CGGTGCGCAA GCTCAGCTAG 1470

The *hemAT-Hs* gene encodes a protein which has an amino acid sequence as shown in SEQ. ID. No. 2 as follows:

MSNDNDTLVTADVRNGIDGHALADRIGLDEAEIAWRLSFTGIDDDTMAALAEQPLFEAT 60
 ADALVDTDFYDHLESYERTQDLFANSTKTVEQLKETQAEYLLGLGRGEYDTEYAQRARIG 120
 KIHVDVLGLGPDVYLGAITYRYTGLLDALADDVVADRGEAAAAVDELVARFLPMLKLLTF 180
 15 DQQIAMDTYIDSYAQRLHDEIDSRQELANAVATHVEAPLSLEATSQDVAERTDTMRART 240
 DDQVDRMADVSRREISSVSASVEEVASTADDVRRTSDEAEALAQGEAAADDALATMTDID 300
 EATDGVTAGVEQLGERAADVESVTGVIDDIAEQTNMLALNASIEAARAGEAGEGFAVVAD 360
 EVKALAEESREQSTRVEELVEQMQAETBETVDQLDEVNQRIEGEVERVEEAMETLQEIIT 420
 AVEDAASGMQEVSTATDEQAVSTEEVAEMVDGVDDRAGEIAAALDDIADATDQQVRTVEE 480
 20 VRETVGKLS 489

In another embodiment of the invention, the heme-binding protein is isolated from *Bacillus subtilis*. Preferably, the *Bacillus subtilis* gene is *hemAT-Bs*, which has a nucleic acid sequence according to SEQ. ID. No. 3, as follows:

ATGTTATTTA AAAAAGACAG AAAACAAGAA ACAGCTTACT TTTCAGATTC AAACGGACAA 60
 CAAAAAACC GCATTACGCT CACAACAAA CATGCAGATG TCAAAAAACA GCTCAAAATG 120
 GTCAGGTTGG GAGATGCTGA GCTTTATGTG TTAGAGCAGC TTCAGCCACT CATTCAAGAA 180
 AATATCGTAA ATATCGTCGA TCGGTTTTAT AAAACCTTG ACCATGAAGC CTCATTGATG 240
 30 GATATCATTA ATGATCACAG CTCAGTTGAC CGCTTAAAC AAACGTAAA ACGGCATATT 300
 CAGGAAATGT TTGACGGCGT TATCGATGAT GAATTTATTG AAAAGCGTAA CCGAATCGCC 360
 TCCATCCATT TAAGAATCGG CTTTTTGCCA AAATGGTATA TGGGTGCGTT TCAAGAGCTC 420
 CTTTGTGCAA TGATTGACAT TTATGAAGCG TCCATTACAA ATCAGCAAGA ACTGTCAAAA 480
 GCCATTAAAG CAACAACAAA AATCTTGAAC TTAGAACAGC AGCTTGTCTT TGAAGCGTTT 540
 35 CAAAGCGAGT ACAACGAGC CGGTGATGAA CAAGAAGAAA AGAAAAACCT TCTTCATCAG 600
 AAAATTCAAG AAACCTCTGG ATCGATTGCC ATTCTGTTTT CAGAAACAAG CAGATCAGTT 660

CAAGAGCTTG TGGACAAATC TGAAGGCATT TCTCAAGCAT CCAAGCCGG CACTGTAACA 720
TCCAGCACTG TTGAAGAAAA GTCGATCGGC GGAAGAAAA AGCTAGAAAT CCAGCAAAAA 780
CAGATGAACA AAATGACAC AAGCCTTGTC CAAATCGAAA AAGAAATGGT CAAGCTGGAT 840
GAAATCGCGC AGCAAATGTA AAAAATCTTC GGCATCGTCA CAGGCATAGC TGAACAAACA 900
5 AACCTTCTCT CGCTCAATGC ATCTATTGAA TCCGCCCGCG CCGGAGAACA CGGCAAAAGC 960
TTTGTCTGCG TGGCAAATGA AGTGCAGGAG CTTTCTGAGG ATACGAAAAA AACCGTCTCT 1020
ACTGTTTCTG AGCTTGTGAA CAATACGAAT ACACAAATCA ACATTGTATC CAAGCATATC 1080
AAAGACGTGA ATGAGCTAGT CAGCGAAAGT AAAGAAAAAA TGACGCAAT TAACCGCTTA 1140
TTCGATGAAA TCGTCCACAG CATGAAAATC AGCAAAGAGC AATCAGGCAA AATCGACGTC 1200
10 GATCTGCAAG CTTTCTITGG AGGGCTTCAG GAAGTCAGCC GCGCCGTTTC CCATGTGGCC 1260
GCTCCGTTG ATTGCTGTGT CATCCTGACA GAAGAATAAC CATCAAAAC CGGTCTGCCA 1320
TACGCCGCGT TTTTTCGCGT TCATTATGTA AACTTAAATT AAAAATCAGT TGACATAATA 1380
ATTACCTGCA 1390

15 In a preferred embodiment, the protein has an amino acid sequence of SEQ.
ID. No. 4, as follows:

MLFKKDRKQETAYFSDSNGQQKNRIQLTNKHADVKKQLKMWRLGDAELVVLBQLQLPIQE 60
NIVNIVDAFYKNLDHESLMDIINDHSSVDRLKQTLKRHIQEMFAGVIDDEFIEKNRRIA 120
20 SIHLRIGLLPKWYMGAFQELLMSMIDIYBASITNQOELLKAIKATTKILNLEQQLVLEAF 180
QSEYNQTRDEQEEKNNLLHQKIQETSGSIANLFSETSRSVQELVDKSEGISQASKAGTVT 240
SSTVEEKSIGGKKELEVQQQKMNKIDTSLVQIEKEMVKLDEIAQQIEKIFGIVTGIAEQT 300
NLLSLNASIESARAGEHGKGFVAVVANEVRKLS EDTKKT VSTVSELVNNTNTQINIVSKHI 360
KDVNELVESKEKMTQINRLFDEIVHSMKISKEQSGKIDVDLQAFGLGQEVSRVSHVA 420
25 ASVDSLVLITTE 432

The invention also provides fragments of the isolated heme-binding protein which contain a functional heme-binding domain. The fragment containing the functional heme-binding domain may be coupled to a heterologous signal transduction domain. As described in the examples, a minimum heme binding domain has been determined for HemAT-*Hs* and partially determined for HemAT-*Bs*. Furthermore, comparisons

between various globin proteins has allowed for the identification of conserved regions of the proteins.

HemAT-*Hs* in *Halobacterium salinarum* and HemAT-*Bs* in *Bacillus subtilis*, the first aerotactic transducers discovered that directly bind oxygen, are heme-based, and are homologous to native sperm whale myoglobin (SWMb), albeit more structural than sequential. These proteins belong to the globin family. Globins bind, transport, and store oxygen, and are known to exhibit a distinctive fold of seven α -helices that encompass a heme prosthetic group. The seven helices are labeled A, B, C, E, F, G, and H. Sometimes, an additional short helix (helix D) is found between helices C and E, as in the case of SWMb, to make a total of eight. In a 1987 publication, Bashford et al., "Determinants of a Protein Fold: Unique Features of the Globin Amino Acid Sequences," J. Mol. Biol., 196:199-216 (1997), which is hereby incorporated by reference, reported that the sequence homology of all the 226 globin sequences known at that time were "as high as 80% or more for closely related species, or as low as 16% for more distant ones." Of all these proteins, only two residues were absolutely conserved throughout. These two residues were the phenylalanine at the end of the C helix (CD1) and the proximal histidine (F8). HemAT-*Hs* and HemAT-*Bs* both contain these two key residues and are 23% and 11% homologous to SWMb, respectively, and share 20% sequence similarity between themselves.

The report of myoglobin-type aerotaxis proteins in microorganisms, and the recent discovery of HemAT-*Hs* and HemAT-*Bs* has prompted an effort to find one or more signature motifs in these possible microbial globins. These would identify conserved regions of the proteins. In addition, with these motifs in hand, contemporary computer algorithms like those contained in the BLAST programs (<http://www.ncbi.nlm.nih.gov/BLAST/>) could permit convenient and rapid searches for other possible globins using this signature motif. These motifs could be used for classifying these newly discovered microbial globins together and eventually with the whole globin family.

Vianogradov et al., "Adventitious Variability? The Amino Acid Sequences of Nonvertebrate Globins," Comp. Biochem. Physiol., 106B:1-26 (1993), which is hereby incorporated by reference, have noted the extensive variation of invertebrate globins over the vertebrates and Bashford et al., "Determinants of a Protein Fold: Unique Features of the Globin Amino Acid Sequences," J. Mol. Biol., 196:199-216 (1987), which is hereby

incorporated by reference, have recognized that alignments of invertebrate globins with vertebrate globins based strictly on sequence similarity and vertebrate data sets are questionable. Invertebrate myoglobins were therefore not included in the preliminary data and the search for a globin motif was limited to vertebrates. Microbial globins, however, were later included and incorporated into the alignment by conserving secondary structure and avoiding gaps as in the work of Kapp et al., "Alignment of 700 Globin Sequences: Extent of Amino Acid Substitution and Its Correlation With Variation in Volume," Pro. Sci., 4:2179-2190 (1995), which is hereby incorporated by reference.

An 80-aa consensus peptide sequence was constructed using the manual alignment of sperm whale myoglobin (SWMb), the oxygen sensor in *Bacillus subtilis*, HemAT-Bs, and the oxygen sensor in *Halobacterium salinarum*, HemAT-Hs. The intent was to find a minimal length of protein containing the myoglobin signature motif and see how many myoglobin proteins this sequence would recognize on the non-redundant (nr) database at NIH using the BLAST server (<http://www.ncbi.nlm.nih.gov/BLAST/>). An X was issued to residues of high variability (Bashford et al., "Determinants of a Protein Fold: Unique Features of the Globin Amino Acid Sequences," J. Mol. Biol., 196:199-216 (1987), which is hereby incorporated by reference) while conserved residues retained their specific amino acid designation. Critical to the alignment was the positioning of the two residues known to be absolutely conserved in all known globins: Phe at the CD1 position and the proximal His at the F8 position (Bashford et al., "Determinants of a Protein Fold: Unique Features of the Globin Amino Acid Sequences," J. Mol. Biol., 196:199-216 (1987), which is hereby incorporated by reference). Using these residues as markers, the myoglobin-like protein (MbLP) sequence was generated and consisted of two domains separated by 32 variable amino acids. The first myoglobin-type domain (M1-box) contained the absolutely conserved phenylalanine residue; the second (M2-box) contained the absolutely conserved proximal histidine. A BLAST search was then performed, comparing the sequences of MbLP and SWMb with those of all other proteins in the non-redundant database. Search parameters were default except for the EXPECT parameter, which was increased to 1000 to allow for matches of lesser sequence homology. This comparison between the number and type of SWMb hits and MbLP hits was used to assess the quality of the MbLP sequence in extracting myoglobin proteins.

A microbial globin-type sequence was generated from the results of a previous BLAST search on microbial globins and included *Vitreoscilla* hemoglobin for structural

markers. This sequence was used to extract 9 bacterial and 8 eukaryotic hemoglobins and flavohemoproteins. This sequence was generated to incorporate microbial globins into the search of a combined globin motif. Manipulation and alignment of the microbial globin-type peptide with MbLP and incorporating the same marker residues produced a second consensus sequence 96-amino acids in length called the triplet globin motif (TGM) because it consisted of three domains: two myoglobin-type domains (M1-box, M2-box) and one hemoglobin-type domain (H-box). TGM was the final sequence used for further analysis and BLASTP searches with the TGM sequence were performed at a lower EXPECT parameter of 600 to reduce the amount of false-positives.

The ability of the myoglobin motif to recognize myoglobins was tested using SWMb as a reference. A BLASTP search of the non-redundant protein database was performed using the 153-aa native sperm whale myoglobin (SWMb) as the query sequence. This sequence recognized 83 unique myoglobins and a wealth of hemoglobins. With some manipulation of the search conditions, however, SWMb was able to extract HemAT-*Hs* as well.

A first attempt at a globin-type motif produced the 80-aa myoglobin-like protein (MbLP) sequence consisting of two domains, the M1-box and M2-box, as found in Figure 3. These two domains recognized 73 myoglobins, or 88% of those found by SWMb, along with HemAT-*Hs*, HemAT-*Bs*, and a few non-globins. In contrast, however, MbLP didn't recognize any hemoglobins.

An effort was made to enhance the globin-type motif of MbLP by building upon itself. This effort resulted in the 96-aa triplet globin motif (TGM) protein sequence and consisted of three domains: the M1-, M2-box, and a new H-box situated in front of the two. The TGM sequence was compared to the MbLP and SWMb by subjecting it to the same BLASTP search analysis. TGM recognized 75 myoglobins (90% of SWMb hits), 17 hemoglobin and hemoglobin-like proteins, and the two HemATs. The 17 hemoglobin and hemoglobin-like proteins consisted of 5 non-microbial eukaryotic hemoglobins from three different organisms and 12 microbial hemoglobins, three eukaryotic and nine bacterial. It is evident that the TGM sequence is more general than MbLP in recognizing globin motifs.

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Based upon the extensive information available regarding conserved structures in the proteins, as well as the minimal functional regions, one can predict modifications to the proteins which will not alter the function of the protein.

The present invention also provides a blood substitute. An urgent need exists among the medical community for an alternative to whole blood or red blood cells for use in transfusion. However, the possibility of transmitting viral infections is ever present in derivatives of human blood. The rapid spread of the AIDS virus as well as the discovery of multiple forms of the virus amplifies this concern. Both HemAT-*Hs* and/or HemAT-*Bs* may present an alternative to whole blood in transfusion situations. HemAT-*Hs* and HemAT-*Bs* are particularly attractive in this regard, since they appear to have low oxygen affinity, a property required for artificial oxygen carriers. Expressed in microorganisms, these oxygen carriers will be free of infectious agents. In addition, the ability of HemAT-*Hs* and HemAT-*Bs* to bind oxygen reversibly and to regulate this binding through the signal transduction domain may lead to new blood substitute products. Currently cross-linked hemoglobins are being developed as blood substitutes but these proteins suffer from poor regulation of oxygen binding. It is possible to develop HemAT-*Hs* and HemAT-*Bs* as a "blood-substitute" due to their size (~50 kDa, similar to hemoglobin) which prevents filtering by the kidney's, its ability to reversibly bind oxygen, and its ability to regulate this binding. Genetically engineered fragments of the *hemAT-Hs* and *hemAT-Bs* genes that encode of the transduction domain provides a wealth of opportunities to regulate oxygen binding.

In a preferred embodiment, the blood substitute has a heme binding domain of the isolated heme-binding protein. The blood substitute may also have a heterologous signal transduction domain, to alter the affinity for oxygen or other gases.

The blood substitute may be administered to a patient suffering from low blood levels. Such a blood substitute has numerous advantages because it could be used as a substitute when whole blood is not available. Furthermore, the blood substitute can be produced so that it is free of infectious substances, such as viruses and bacteria.

In addition to using heterologous signaling domains, the oxygen binding of the heme-binding protein may be altered by modifying the signaling domain.

The invention also provides a method for controlled storage of oxygen. The bacterial heme binding protein can be contacted with oxygen allowing the protein to bind and store oxygen. The protein may also be covalently attached to a solid substrate via the

transduction domain. Subsequent triggering of the transduction domain can result in oxygen release.

The present invention may also be used to sense gaseous ligands by exposing the bacterial heme binding protein to a sample to be tested and measuring a change in the conformation of the protein. Enzyme sensors are well known as biological sensors. They are utilized mainly for clinical chemical analysis, including use for glucose in blood, urea and neutral and phospholipids. The ability of HemAT-*Hs* and HemAT-*Bs* to sense oxygen as well as other small gaseous ligands provides opportunities to develop novel biosensors for O₂, NO, CO, and even CN⁻.

The changes in the conformation of the protein may be monitored in various ways including monitoring the protein optically or electronically.

The preferred gaseous ligands to monitor with the heme binding bacterial protein are O₂, NO, CO, and CN⁻. The preferred gaseous ligand is O₂.

As discussed above fragments of the bacterial heme binding protein may also be used as long as they contain a functional heme binding domain.

The present invention also provides a chimeric protein having a heme-binding domain of an isolated heme binding bacterial protein and a heterologous signaling domain. Varying the signaling domain can alter the oxygen or ligand binding characteristics of the protein. The signaling domain may also be altered to make the protein responsive to other signals.

In another embodiment, the invention provides an isolated nucleic acid molecule which encodes a bacterial heme binding protein with a heterologous or mutated signaling domain.

The bacterial heme-binding proteins may also be used for heme-based catalysis.

It is well known that Fe(III)porphyrins can catalyze a wide variety of chemical reactions including hydrogen peroxide degradation, mono oxygenation, and lignin degradation. HemAT-*Hs* can also be prepared in the Fe(III) form providing an opportunity to utilize this protein as a novel heme-based catalyst. In addition, the ability to regulate the heme domain by the transduction domain may allow for catalytic specificity to be achieved via genetic manipulation of this domain.

The proteins of the present invention may also be used for artificial photosynthesis. HemAT-*Hs* can be reconstituted with different porphyrins including photoactive Zn and Sn derivatives. These derivatives may possess the ability to absorb

light energy and transmit information concerning the excited state of the photoactive porphyrin to the sensing domain providing the equivalent to photosynthesis, i.e., conversion of light energy to chemical potential energy.

The bacterial heme binding proteins may also be used *in vivo* and *in vitro* testing system for identifying potential signaling functions of mutated α -hemoglobin and myoglobin causing several diseases. Mutated human α -hemoglobin and myoglobin genes can be fused with fragment of *hemAT-Hs* or *hemAT-Bs* genes that encodes signaling domain via linker region. The physiological function of the expressed chimeric protein of human α -hemoglobin (or myoglobin) and HemAT-*Hs* or HemAT-*Bs* can be tested by capillary aerotaxis assay. As transducer proteins HemAT-*Hs* and HemAT-*Bs* may cause phosphorylation of CheA. Once this feature of HemAT-*Hs* and HemAT-*Bs* are tested and optimized, similar *in vivo* strategy of chimeric protein construction can be tested for *in vitro* phosphorylation assay.

EXAMPLES

Example 1 – Mutagenesis of HemAT-*Hs* and HemAT-*Bs*

The HtrVIII is a positive aerotaxis transducer in *H. salinarum* (Brooun et al., J. Bacteriol., 180:1642-1646 (1998), which is hereby incorporated by reference). A strain deleted for the *htrVIII* gene lacks positive aerotaxis while a strain overproducing the protein shows an enhanced aerotactic response. To investigate the possible role of HemAT-*Hs* and HemAT-*Bs* in aerotaxis, deletion mutants of these genes were constructed (Brooun, Ph.D thesis. University of Hawaii, Hawaii (1997), which is hereby incorporated by reference) for the construction of *hemAT-Hs* deletion strains. Construction of overexpression of *hemAT-Hs* in *H. salinarum*: *NdeI* and *XbaI* restriction sites were used to clone the *hemAT-Hs* gene into the *E. coli-H. salinarum* shuttle vector pKJ427. Top primer with *NdeI* cutting site (5'CCGAATTCATATGAGCAACGAT AATGAC 3' (SEQ. ID. No. 40)) and bottom primer with *XbaI* cutting site (5'CCTCTA GAGGATEECTAGCTGAGCTTGCCGACC 3' (SEQ. ID. No. 41)) were synthesized and used for PCR amplification of *hemAT-Hs* gene. The PCR amplicon was cloned into TOPO cloning vector (Invitrogen) and transformed into *E. coli* competent cells. The plasmid containing *hemAT-Hs* gene in TOPO vector was subcloned into pKJ427 vector

by *NdeI/XbaI* double digestion. The *hemAT-Hs/pKJ427* construction was confirmed by PCR as well as *NdeI/XbaI* double digestion and transformed into $\Delta htrVIII$ strain using standard halobacteria transformation protocol. Individual colonies were checked by PCR and immunoblot to confirm the expression level of HemAT-*Hs*; Construction of OI3428:

- 5 A 322 bp fragment interior to HemAT-*Bs* was amplified from the *B. subtilis* wild type strain OI1085 chromosome using primers with overhanging *HindIII* and *BamHI* sites (reverse primer: 5' TATGGGATCCCTTGTCATCACGGGTCTETTGG 3' (SEQ. ID. No. 42), forward primer: 5' GATAAAGCTTGATCATAGCTCAGTTGACCG 3' (SEQ. ID. No. 43)). This PCR fragment was digested with *HindIII* and *BamHI* and
- 10 cloned in the integration vector pHV501 (Vagner et al., Microbiology, 144(Pt 11):3097-3104 (1998)) to create pMK1. The resultant plasmid pMK1 was transformed into OI1085 and HemAT-*Bs* mutants were selected by erythromycin resistance. Integration of the pMK1 into the correct locus was checked by linkage analysis. The *hemAT-Bs* locus is 30% linked to the *glyk* locus as determined from the *B. subtilis* chromosomal map.
- 15 GLY+ transductants were selected and scored for erythromycin resistance. Construction of OI3498: The entire HemAT-*Bs* gene including the native promoter and the ribosome binding site was amplified from the *B. subtilis* wild type strain OI1085 chromosome using primers with overhanging *EcoRI* and *BamHI* sites (HemAT-*Bs* amyup: 5' TGCTGAATTCGAGCTTTCATTCATGTTTCCC 3' (SEQ. ID. No. 44), HemAT-*Bs* amydown: 5' TTAGGGATCCGTCAACTGATTTTAA TTAAAGTTAC 3') (SEQ. ID. No. 45)). The PCR amplicon was digested with *EcoRI/BamHI* and cloned into the amyE integration vector pDG1730 (Guerout-Fleury et al., Gene, 180(1-2):57-61 (1996), which is hereby incorporated by reference) to produce pKZ2. The resultant plasmid pKZ2 was digested with *BglI/XbaI* to ensure a double crossover event into the amyE
- 20 locus and then transformed into OI3428 to select for Spec-R. HemAT-*Bs* overexpression R4: Overexpression construction in *E. coli*: The HemAT-*Bs* overexpression construction was performed as follows: *B. subtilis* OI1085 genomic DNA was used for the PCR amplification of HemAT-*Bs* gene by Pfu DNA polymerase using two primers (Top primer with *BamHI* restriction site: 5'ATATGGATCC
- 25 AAGGGGGATCATTGTAATGTTATTTAAAAAAG3' (SEQ. ID. No. 46), Bottom primer with *PstI* site: 5' ATTACTGCAGCAACTGATTTTAAATTTAAGTTT ACATAATGAACGC 3' (SEQ. ID. No. 47)). The PCR amplicon was cloned into TOPO cloning vector (Invitrogen) and transformed into TOP 10 *E. coli* competent cells.
- 30

Colonies were tested for the presence of plasmids containing the correct insert. The recombinant plasmid was digested with *Bam*HI and *Pst*RI and the insert with HemAT-*Bs* open reading frame was cloned into the pMALcII expression vector (New England Biolabs, Inc).

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Example 2 – Aerophilic and Aerophobic Responses

The resultant construct was transformed to *E. coli* pLysS cells for the expression and analyzed their behavior in a flat microcapillary using dark-field microscope coupled with time-lapse digital video system. Motile wild-type halobacterial cells form two clear congregated aerotactic bands, a positive one close to the interface between air and cell suspensions and a negative one away from the interface (Figure 5A, wild type). The positive aerotactic band is mediated by HtrVIII (Brooun et al., *J. Bacteriol.*, 180:1642-1646 (1998), which is hereby incorporated by reference). As expected, this phenomenon is absent in the *htrVIII* deletion strain (Figure 5A, HemAT-*Hs*+ΔHtrVIII). However, like the wild type strain, the ΔHtrVIII strain also demonstrates the negative aerotactic band. If negative aerotaxis behavior is related to HemAT-*Hs*, one would postulate that in the *hemAT-Hs* deletion strain, the negative aerotactic band would not form. Indeed, in the Δ*hemAT-Hs* strain, in which the positive aerotactic band is present due to the receptor HtrVIII, the sharp boundary of the negative aerotactic band is absent (Figure 5A, Δ*hemAT-Hs*). Furthermore, when HemAT-*Hs* is overexpressed (using a multicopy plasmid) in a Δ*htrVIII* strain, halobacterial cells form a more pronounced negative aerotaxis boundary (Figure 5A, HemAT-*Hs*+ΔHtrVIII). These cells were repelled from the air/liquid interface much faster and created a denser aerotactic band than the aerotactically wild type or ΔHtrVIII strains containing genomic copy of *hemAT-Hs* (Figure 5A, wild type and HemAT-*Hs*+ΔHtrVIII). The aerophilic response in *B. subtilis* proceeds more rapidly than it does in *H. salinarum* (30 versus 180 min) because *B. subtilis* swims faster than *H. salinarum*. In the wild-type, an aerotactic band formed at the air interface (Fig. 5B). This band did not form in a strain from which all ten putative MCP-like transducers (Δ*ten*) were deleted (Fig. 5B). A strain lacking only HemAT-*Bs* showed an aerophobic response, indicating the presence of a second, unidentified aerotaxis receptor. To demonstrate the physiological function of HemAT-*Bs*

unequivocally, *hemAT-Bs* was overexpressed in a strain from which all *B. subtilis* transducer genes were deleted (Δ ten strain). When HemAT-*Bs* was overexpressed in the Δ ten strain, the aerophilic response was observed (Fig. 5b). These assays demonstrate that HemAT-*Bs* is involved in an aerophilic response in *B. subtilis*.

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Example 3 – Expression of HemAT-*Hs* and HemAT-*Bs* in *Escherichia coli*

The FAD-binding aerotaxis transducer Aer in *E. coli* has a PAS domain that is similar to the redox-sensing domain of the NifL protein of *Azotobacter vinelandii* (Hill et al., Proc. Natl. Acad. Sci. USA, 93:2143-2148 (1996); Zhulin et al., Mol. Microbiol., 29:1522-1523 (1998), which are hereby incorporated by reference) and FixL from *R. meliloti* (Gilles-Gonzalez et al., Nature, 350:170-172 (1991), which is hereby incorporated by reference). FixL is a chimeric membrane protein with a histidine kinase domain, which belongs to the large class of two-component regulatory systems, whereas the heme-binding sensory domain belongs to the PAS domain super family (Gilles-Gonzalez et al., Nature, 350:170-172 (1991); Lois et al., J. Bacteriol., 175:1103-1109 (1993); Gong et al., Proc. Natl. Acad. Sci. USA, 95:15177-15182 (1998), which are hereby incorporated by reference). None of the PAS domains identified in the genome of *B. subtilis* is present in chemotaxis transducers (Zhulin et al., Mol. Microbiol., 29:1522-23 (1998), which is hereby incorporated by reference). To identify the nature of the prosthetic groups in HemAT-*Hs* and HemAT-*Bs*, both proteins were expressed in *E. coli* by constructing vectors, which express the *hemAT-Hs* or *hemAT-Bs* gene under the control of an inducible T7 promoter (Studier et al., Methods in Enzymology, 185:60-89 (1990), which is hereby incorporated by reference).

Using a combination of anion exchange and gel-filtration chromatography, HemAT-*Hs* was purified (The BL21 pLysS host cells harboring *hemAT-Hs* or *hemAT-Bs* genes were grown to $OD_{600} = 0.4$ in 1L of LB with appropriate antibiotics and induced with 0.6 mM IPTG. The cells were harvested by low speed centrifugation (4000xg) for 15 min. at 4°C after a two-hour induction. The pellets were resuspended in 50 ml buffer (50 mM NaCl, 50 mM Tris-HCl, pH6.0) and sonicated for a total of 4 minutes (20 second pulses with 30 second pauses). The sonicated solution was centrifuged at 28,000xg for 20 min. The brown red supernatant with HemAT-*Hs* or HemAT-*Bs* was used for purification. **HemAT-*Hs*:** The supernatant was filtered through 0.2 micron filter and

applied to BioCAD anion exchange POROS HQ/M (16/100) perfusion chromatography column equilibrated with 50 mM Tris-HCl, pH6.0. A linear gradient of NaCl (0-1500 mM) was applied and HemAT-*Hs* was eluted at about 400 mM. For further purification, the fractions containing the HemAT-*Hs* (monitored by Soret band absorbance at 410 nm and SDS-gel electrophoresis) were concentrated and applied to a Hiload Superdex 200 16/60 gel filtration column. Peak fractions were concentrated with an Amicon 100K concentrator and used for spectroscopy. **HemAT-*Bs*:** A saturated (NH₄)₂SO₄ solution was added to the brown red supernatant to 30% and centrifuged at 28,000xg for 20 min. The optically clear light brown supernatant was further fractionated by (NH₄)₂SO₄ addition to 36% saturation followed by centrifugation. The resultant pellet was solubilized in a resuspension buffer (500mM NaCl, 50mM Tris-HCl, pH8) and applied to a Hiload 26/60 Superdex 75 gel filtration column. Peak fractions containing HemAT-*Bs* (monitored by Soret band absorbance at 410 nm and SDS-gel electrophoresis) were concentrated by an Amicon 50K concentrator and used for spectroscopy). Recombinant HemAT-*Hs* expressed in *E. coli* under low ionic strength conditions was shown to contain a high degree of secondary structure consistent with a predicted folded protein (Larsen et al., J. Prot. Chem., 18(3) (1999), which is hereby incorporated by reference).

The purified HemAT-*Hs* migrates at a position higher than the calculated 52.8 kDa for the mature protein (Figure 2B line HemAT-*Hs*). This slow electrophoretic migration in SDS-polyacrylamide gels is consistent with the highly acidic nature of HemAT-*Hs* (pI=3.78, 27% acidic residues) and has been observed in other acidic proteins from halophiles (Ihara et al., Arch. Biochem. Biophys., 286:111-116 (1991), which is hereby incorporated by reference). Using a combination of ammonium sulfate precipitation/fractionation and gel filtration chromatography it is possible to purify HemAT-*Bs*. The BL21 pLysS host cells harboring *hemAT-Hs* or *hemAT-Bs* genes were grown to OD₆₀₀ = 0.4 in 1L of LB with appropriate antibiotics and induced with 0.6 mM IPTG. The cells were harvested by low speed centrifugation (4000xg) for 15 min. at 4°C after a two-hour induction. The pellets were resuspended in 50 ml buffer (50 mM NaCl, 50 mM Tris-HCl, pH6.0) and sonicated for a total of 4 minutes (20 second pulses with 30 second pauses). The sonicated solution was centrifuged at 28,000xg for 20 min. The brown red supernatant with HemAT-*Hs* or HemAT-*Bs* was used for purification.

HemAT-*Hs*: The supernatant was filtered through 0.2 micron filter and applied to BioCAD anion exchange POROS HQ/M (16/100) perfusion chromatography column

equilibrated with 50 mM Tris-HCl, pH6.0. A linear gradient of NaCl (0-1500 mM) was applied and HemAT-*Hs* was eluted at about 400 mM. For further purification, the fractions containing the HemAT-*Hs* (monitored by Soret band absorbance at 410 nm and SDS-gel electrophoresis) were concentrated and applied to a Hiload Superdex 200 16/60 gel filtration column. Peak fractions were concentrated with an Amicon 100K concentrator and used for spectroscopy. **HemAT-*Bs*:** A saturated $(\text{NH}_4)_2\text{SO}_4$ solution was added to the brown red supernatant to 30% and centrifuged at 28,000xg for 20 min. The optically clear light brown supernatant was further fractionated by $(\text{NH}_4)_2\text{SO}_4$ addition to 36% saturation followed by centrifugation. The resultant pellet was solubilized in a resuspension buffer (500mM NaCl, 50mM Tris-HCl, pH8) and applied to a Hiload 26/60 Superdex 75 gel filtration column. Peak fractions containing HemAT-*Bs* (monitored by Soret band absorbance at 410 nm and SDS-gel electrophoresis) were concentrated by an Amicon 50K concentrator and used for spectroscopy). The purified HemAT-*Bs* migrates in SDS-PAGE as 48.7 kDa protein as expected (Figure 2, line HemAT-*Bs*).

Example 4 – Absorption Spectra of Purified HemAT-*Hs* and HemAT-*Bs*

HemAT-*Hs* and HemAT-*Bs* display similar absorption spectra in both the near UV and visible regions characteristic of oxygen bound heme proteins. Specifically, absorption band maxima are found at 406 nm (Soret), 578 nm (α -band), and 538 nm (β -band) for both proteins (Figure 4A). These absorption maxima resemble those of *Sperm* whale oxymyoglobin (418 nm, 581 nm, and 543 nm) and oxy FixL (415 nm, 577 nm, and 543 nm). Upon deoxygenation (using sodium dithionite), the Soret bands shift to 425 nm while the α - and β -bands converge to a broad band centered at 555 nm, consistent with the formation of a deoxy-form of the protein (i.e., absorption bands for deoxymyoglobin: 434 nm and 556 nm and deoxyFixL: 433 nm and 567 nm) (Figure 4B). If the deoxy form of HemAT-*Hs* and HemAT-*Bs* are exposed to atmospheric oxygen, the absorption spectra revert back to that observed for the purified proteins (Figure 4D). Both the purified (oxy form) and the deoxy derivatives of HemAT-*Hs* and HemAT-*Bs* are reactive towards carbon monoxide. The CO bound derivatives display absorption maxima at 415 nm (Soret), 573 nm (α -band), and 535 nm (β -band) (Figure 4C). A pyridine hemochrome

assay showed the heme group of both HemAT-*Hs* and HemAT-*Bs* to be b-type. HemAT-*Hs* and HemAT-*Bs* are distinct both in spectral features and in physiological function from the previously discovered heme protein FixL from *R. meliloti* (Gilles-Gonzalez et al., Nature, 350:170-172 (1991), which is hereby incorporated by reference). The absorption bands of both HemAT-*Hs* and HemAT-*Bs* are blue shifted, relative to FixL, indicating distinct heme pocket geometries. Unlike FixL, HemAT-*Hs* and HemAT-*Bs* display no PAS domain sequence homology. In addition, both HemAT-*Hs* and HemAT-*Bs* participate in negative aerotaxis while FixL acts as an oxygen sensing kinase.

10 **Example 5 - Methylation of HemAT-*Hs* and HemAT-*Bs***

It has been postulated that in *E. coli*, adaptation in Aer-mediated aerotaxis is methylation-independent (Taylor et al., Annu. Rev. Microbiol., 53:90-103 (1999), which is hereby incorporated by reference). In contrast to *E. coli*, adaptation during aerotaxis in *H. salinarum* and *B. subtilis* is a methylation-dependent process (Brooun et al., J. Bacteriol., 180:1642-1646 (1998); Lindbeck et al., Microbiology, 141:2945-2953 (1995); Wong et al., J. Bacteriol., 177:3985-3991 (1995), which are hereby incorporated by reference). To determine if HemATs can be methylated by the CheR methyltransferase, *H. salinarum* and *B. subtilis* cells were radiolabeled with [*methyl-³H*] methionine after blocking protein synthesis. The radiolabeled cells were processed for fluorography and immunoblotting with a polyclonal antibody raised against the highly conserved region of methyl-accepting transducers (W. Zhang et al., Proc. Natl. Acad. Sci. U.S.A., 93:4649 (1996), which is hereby incorporated by reference). A single radiolabeled band is missing in the Δ *hemAT-Hs* strain (Figure 2B, lane 1), whereas this band is present in the overexpression strain (Figure 2B, lane 2). This band is also recognized by the antibody, suggesting that HemAT-*Hs* is indeed a methyl-accepting transducer (Figure 2B, lanes 1' and 2'). In contrast, it was not possible to detect any [*methyl-³H*] labeling in HemAT-*Bs*. Together with the capillary assays, these data demonstrate an important difference in the signaling and adaptation mechanisms for aerotaxis mediated by HemAT-*Hs* and HemAT-*Bs*.

Example 6 – Materials and Methods for Example 7

PCR and TOPO cloning. C-terminal primers were designed to amplify the
5 250,230,210, 205, 200, 195, 194, 193, 192, 191, 190, 170 and 151 residues of HemAT-*Hs*
and were designed to include *Bam*HI and *Xba*I restriction sites. The N-terminal primer
included *Eco*RI and *Nde*I restriction sites. Primer sequences may be found in Table 1.
HemAT-*Hs* genomic plasmid was used as a template for PCR with *Pfu* polymerase. PCR
amplification was performed in a GeneAmp PCR system 2400 (Perkin-Elmer) under the
10 following conditions: Hot start with *Pfu* polymerase at 80°C followed by heat
denaturation at 94°C for 2 minutes was followed by 25 cycles of heat denaturation at 94°C
for 30 seconds, primer annealing at 58°C for 30 seconds and elongation at 72°C for 40
seconds. Following the last cycle, samples were maintained at 72°C for 7 minutes and
immediately kept at 4°C. Following PCR, the PCR product was immediately cloned into
15 the TOPO vector (Invitrogen) and transformed into TOP10 competent cells. Clones with
the insertion were selected via kanamycin resistance on Luria Bertani (LB) agar plates
with kanamycin (50µg/ml). Colonies were inoculated into CircleGrow (BIO101) with
kanamycin media and, following incubation, plasmids were isolated via alkaline mini
prep. Plasmids were then restricted with *Eco*RI to screen for the proper insert.

20 **Cloning into pMAL expression vector.** Plasmids containing the correct insert
and the expression vector, pMAL-c2, were then digested with *Eco*RI and *Bam*HI. pMAL-
c2 was subsequently dephosphorylated with Alkaline Phosphatase. Digested TOPO
plasmids and pMAL plasmid were run on a 1% preparative agarose gel. The truncated
hemAT-Hs PCR insert and double digested pMAL-c2 bands were cut from the gel and the
25 DNA was extracted from the gel using the GENECLEAN Spin Kit (BIO101). The
hemAT-Hs insert was then ligated to the pMAL-c2 vector at 14°C, overnight. Following
ligation, the ligation mixture was transformed into JM109 competent cells. Clones
containing the plasmid were selected for by ampicillin resistance on LB agar Amp
(100µg/ml) plates. Ampicillin resistant colonies were inoculated into CircleGrow + Amp
30 media and incubated. Plasmids were isolated via alkaline mini prep and the *hemAT-Hs*
insert was screened for by double digest with *Eco*RI and *Bam*HI.

Transformation into expression host and protein expression. Plasmids
containing the insertion were then transformed into BL21 *pLysS* competent cells

(Novagen). Clones containing both the pMAL-*hemAT-Hs* insertion plasmid and the pLysS plasmid were screened for by ampicillin (100µg/ml) and chloramphenicol (34µg/ml) resistance on LB agar plates. To check for expression of the truncated MBP-HemAT-*Hs* fusion protein, cells were inoculated into LB Amp and Chl broth and grown to an OD₆₀₀=0.4 followed by induction with 1 mg/ml IPTG. After induction for 1.5 hours, protein samples of uninduced and induced cultures were prepared and run on a 10% SDS-PAGE. This was then followed by staining for protein with Coomassie Blue and destaining with 10% acetic acid.

Protein purification by affinity chromatography and spectral analysis.

Cultures which showed induction of the MBP-HemAT-*Hs* protein were then grown up in a larger scale to OD₆₀₀=0.4 and induced with IPTG (1 mg/ml). Induced cultures were then centrifuged at 5,000 rpm for 20 minutes at 4°C followed by a wash with column buffer (20mM Tris-HCl, 200mM NaCl, 1mM EDTA) and centrifuged again at 5,000 rpm for 20 minutes at 4°C. If purification did not immediately follow the wash, protein pellets were stored at -70°C. Protein pellets were then resuspended in column buffer, sonicated for 2 minutes (20 second pulses at 45 second intervals) resuspended in column buffer, sonicated for 2 minutes (20 second pulses at 45 second intervals) and centrifuged at 15,000 rpm for 20 minutes at 4°C. The protein containing supernatant was decanted, diluted 1:2 and stored on ice. After setting up the amylose resin column (New England BioLabs), it was washed with 8 column volumes of cold column buffer. The sample was then loaded onto the column at a flow rate of 1 ml/min. followed by a 12 column volume wash with cold column buffer. MBP-HemAT-*Hs* protein was eluted with 10 mM maltose column buffer and collected in 1 ml fractions. Samples containing the most protein were used to determine the spectra via spectrophotometer. Following elution, a 10% SDS-PAGE was also often run to determine the amount of protein in elutions. Eluted samples were stored at 4°C.

Table 3. Names and sequences (5' to 3') of primers used in HemAT-*Hs* truncation.

Primer Name	Sequence (5' to 3')	
<i>hemAT-Hs EcoRI/NdeI</i> top	ccgaattccatgatgacacgataatgac	SEQ. ID. No. 48
<i>hemAT-Hs 151 BamHI/XbaI</i> bot	ctctagaggatccctagtcgtcggcaagcgcgtcc	SEQ. ID. No. 49
<i>hemAT-Hs 250 B/X</i> bot	ctctagaggatccctagacgtcagccatgcggtc	SEQ. ID. No. 50
<i>hemAT-Hs 230 B/X</i> bot	ctctagaggatccctagcgcagctcctgcgaggtcgcc	SEQ. ID. No. 51
<i>hemAT-Hs 210 B/X</i> bot	ctctagaggatccctacgcgttcgccaactcctgcgcgc	SEQ. ID. No. 52
<i>hemAT-Hs 190 B/X</i> bot	ctctagaggatccctagatgtatgtctcattgcgac	SEQ. ID. No. 53
<i>hemAT-Hs 170 B/X</i> bot	ctctagaggatccctacggccacgagttcgtcgac	SEQ. ID. No. 54
<i>hemAT-Hs 205 B/X</i> bot	ctctagaggatccctactggcgcgtgtcgtatctgic	SEQ. ID. No. 55
<i>hemAT-Hs 200 B/X</i> bot	ctctagaggatccctactcgtcgtggagcgctgggc	SEQ. ID. No. 56
<i>hemAT-Hs 195 B/X</i> bot	ctctagaggatccctactggcgtacgagtcgatgtag	SEQ. ID. No. 57
<i>hemAT-Hs 194 B/X</i> bot	ctctagaggatccctagggctacgagtcgatgtaggtgcc	SEQ. ID. No. 58
<i>hemAT-Hs 193 B/X</i> bot	ctctagaggatccctagtagcagtcgatgtaggtgcc	SEQ. ID. No. 59
<i>hemAT-Hs 192 B/X</i> bot	ctctagaggatccctacgagtcgatgtaggtgcattgcg	SEQ. ID. No. 60
<i>hemAT-Hs 191 B/X</i> bot	ctctagaggatccctagtcgatgtaggtgcattgcg	SEQ. ID. No. 61

Example 7 – Truncated HemAT-*Hs*

The finding that HemAT-*Hs*, an archaeal signal transducer, is a heme binding protein provides a unique opportunity to study not only the physiological function of this protein, but also obtain greater understanding of the structure of this soluble protein and how heme interacts with it. Therefore, this project aims to identify the minimum size of HemAT-*Hs* to which heme binds. This will be done by truncating the gene, first from the C - terminal, by PCR. Once the minimum size of the functional heme binding domain is found from the C - terminal, the N - terminal will then be truncated to further identify residues crucial in proper heme binding. Producing this truncated HemAT-*Hs* protein which still retains the functional heme binding domain will aid in efforts to determine HemAT-*Hs* protein structure.

Analysis of heme binding in HemAT-*Hs* began with the first 151 residues of HemAT-*Hs*. However, preliminary spectral analysis showed no heme binding. Primers were then designed to amplify HemAT-*Hs* every 20 amino acids from 150 thereby amplifying the first 170, 190, 210, 230, and 250 amino acids of the N-terminal. HemAT-*Hs* 210 showed the spectra of heme bound to HemAT-*Hs* and also exhibited the characteristic red spectra of O₂ bound heme in purified protein samples. HemAT-*Hs* 190, however, did not present color in protein samples, nor did it have the visible bands at 540 nm and 580 nm which represent bound heme. Primers were then designed every 5 amino

acids from 210 to 190 at 205, 200 and 195 to determine more precisely where heme binds. Only 200 and 195 construction fused with MBP showed a reddish color in protein samples along with the characteristic spectra.

5 This narrowed the search down for the heme binding site to between 195 and 190 amino acids of HemAT-*Hs*; thus, primers were designed at 191, 192, 193 and 194 amino acids. The spectra for the 192 and 191 constructs shows altered visible bands at 540nm and 580nm. The 194 construction shows a similar spectra, like wild-type HemAT-*Hs* or the HemAT-*Hs* 195 construct.

10

Example 8 – Purification of Recombinant HemAT-*Hs* by Metal Chelate Chromatography

15 1 L of *E. coli* culture containing HemAT-*Hs* was collected, washed with Buffer #2 (200 mM NaCl, 50 mM Sodium phosphate, pH 8.0), and resuspended in 40 ml of Buffer #2. Cells are sonicated. Insoluble material is removed by ultracentrifugation at 100,000 rpm for 20 minutes. POROS MC/M (100 X 1.6 I.D., 20 μ m) is used for metal chelate chromatography. The column is washed with 50 mM EDTA, 1 M NaCl, pH 8.0 over 10 column volumes followed by a wash with water. 100 mM CoCl₂ is used to charge the column, followed with a wash with 1 M NaCl and water. The column is equilibrated in
20 buffer containing 5 mM imidazole. 5 ml of sample is loaded directly onto the column at a flow rate of 2-4 ml/min and a gradient of imidazole from 0-500 mM is run over 30 column volumes at 10 ml/min. Fractions containing recombinant HemAT-*Hs* is pooled and concentrated using Centricon 50.

Example 9 - Site Directed Mutagenesis of HemAT-Hs

In order to perform PCR-based site-directed mutagenesis, a plasmid containing the *hemAT-Hs* gene to be mutated with proper size has to be constructed first. The proper restriction recognition sites are created by designing the primers with the recognition site tags in the primers as follows:

hemAT-Hs *EcoRI/NdeI* top primer:

5' CCGAATTCATATGAGCAACGATAATGAC 3' (SEQ. ID. No. 62)

hemAT-Hs *BamHI/XbaI* bottom primer:

5' CCTCTAGACTAGCTGAGCTTGCCGACC 3' (SEQ. ID. No. 63)

Two sites in each primer were created to meet the needs of expressing HemAT-Hs in different expression vectors. *hemAT-Hs* genomic DNA in pDelta vector was used as a template for amplifying *hemAT-Hs* gene by PCR using proofreading DNA polymerase *pfu*. PCR product was cloned into TOPO vector (Invitrogen TOPO cloning Kit). The insert was checked and confirmed by digestion and PCR. This construction was used as template for generating serial histidine mutants.

The plasmid construction from above was used for mutagenesis PCR. His 20, His 71, His 123, His 198, and His 214 were mutated to alanine by PCR-based site-directed mutagenesis (described above). Mutated *hemAT-Hs* gene in Topo vector has been checked by manual sequence as well as Auto Sequencer 373.

Table 4: Primers for mutagenesis.

Primer Name	Sequence	
H20A	GGAACGGGATCGACGGGgcccGCACTCGCGGACCGG	SEQ. ID. No. 64
H20A-R	CCGGTCCGCGAGTGCgcccCGGTCGATCCCGTTCC	SEQ. ID. No. 65
H70A	GACCGACTTCTACGACgcccTTGGAGTCTTACGAGCG	SEQ. ID. No. 66
H70A-R	CGCTCGTAGGACTCCAAGgcccGTCGTAGAAGTCGGTC	SEQ. ID. No. 67
H123A	CCGTATCGGGAAGATAgcccGACGTGCTCGGGCTCG	SEQ. ID. No. 68
H123A-R	CGAGCCCGAGCAGCTCgcccTATCTTCCCGATACGG	SEQ. ID. No. 69
H198A	CGTACGCCAGCGCCTCgcccGACGAGATCGACAGCC	SEQ. ID. No. 70
H198A-R	GGTGTGATCTCGTGCgcccGAGGCGCTGGGCGTACG	SEQ. ID. No. 71
H214A	GCGAACGCGTCCGACGgcccGTGGAAGCACCGCTG	SEQ. ID. No. 72
H214A-R	CAGCGGTGCTTCCACgcccGTCYGCACCGCTTCGC	SEQ. ID. No. 73

Total of 10 mutants have been done, including H20A, H70A, H123A, H198A, H214A, H20/70A, H20/123A, H70/123A, H20/70/123A.

Example 10 - Expression of mutated *hemAT-Hs*

The *hemAT-Hs*/pTOPO construction was used as initial plasmid for the
5 subcloning of *hemAT-Hs* gene into different vectors. Three different expression systems
were used. First, the pMAL system was used for expression in *E. coli* (Fusion protein
expression system). *EcoRI* and *BamHI* restriction digestion sites were used for cloning
the mutated *hemAT-Hs* gene into pMAL vector. The protein expressed in this system is a
MBP HemAT-*Hs* fusion protein. All of the mutants have been cloned into pMAL,
10 expressed successfully, purified and spectra have been done as well.

Second, the pET system is also used for expression of the peptides in *E. coli*.
NdeI and *BamHI* restriction digestion sites were used for subcloning *hemAT-Hs* into pET
vector.

Third, in order to study the physiological function of HemAT-*Hs* in its native
15 host, it has to be expressed in halobacterial Δ *hemAT-Hs* strain, a strain that *hemAT-Hs*
gene has been deleted from its genome. *NdeI* and *XbaI* were used to clone mutated
hemAT-Hs gene into a halobacterial shuttle expression vector pKJ427. *hemAT-*
Hs/pTOPO plasmid was digested with *NdeI* and *XbaI*, as well as the shuttle vector
pKJ427. Digested vector and *hemAT-Hs* insert were purified from agarose gel by
20 GeneClean kit and ligated with T4 ligase at 4°C. Ligation reaction was transformed into
E. coli competent cells. Colonies were inoculated, the plasmids were extracted and
checked by double digestion and PCR. The final construction was transformed into
halobacterial *hemAT-Hs* deletion strain for over-expressing HemAT-*Hs* in *H. salinarum*
(standard halobacterial transformation protocol was used). Cultures were checked for
25 expression of HemAT-*Hs* by immunoblot using both HC23 antibody and HemAT-*Hs*
specific antibody. The clone with highest expression of HemAT-*Hs* was used for
physiological study.

Example 11 – Construction of a C-terminal His-tag of *hemAT-Hs*

30

In order to purify HemAT-*Hs* protein from its native host *Halobacterium*
salinarum C-terminal His-tag was constructed. A two-step PCR strategy was used. First,
an *NdeI* top primer and 20 nucleotide C-terminal of *hemAT-Hs* gene plus sequence

encoding 6-histidine primer were used for amplification. Second, using first round PCR product as template, *NdeI* top primer and 6-Histidine + Stop codon bottom primer were used for PCR. The primer (including *BamHI/XbaI* cutting sites) was used to amplify the *hemAT-Hs* gene plus histidine codon as well as stop Codon right after 6-his sequence.

- 5 TOPO cloning was used for cloning the PCR products. *NdeI/XbaI* were used for subcloning of *hemAT-Hs*-6-His-stop construction into shuttle vector pKJ427. The final construction plasmid was transformed into *hemAT-Hs* deletion strain.

Example 12 - HemAT-Hs overexpression construction in *H. salinarum*

10

Expression of HemAT-Hs in *Halobacterium salinarum* was created in the expression vector pKJ 427. pKJ 427 plasmid contains a fedox promotor with an mevinolin resistant gene. *NdeI* and *XbaI* restriction recognition sites were used to clone the *hemAT-Hs* gene into the pKJ427 vector. Top primer with *NdeI* cutting site and
15 bottom primer with *XbaI* cutting site were designed and used for amplifying *hemAT-Hs* gene from *Halobacterium salinarum* genomic DNA by proof-reading pfu DNA polymerase. The PCR product was cloned into TOPO cloning vector (Invitrogen) and transformed into *E. coli* competent cells. The plasmid containing *hemAT-Hs* gene in TOPO vector was subcloned into pKJ 427 vector by *NdeI/XbaI* double digestion. The
20 *hemAT-Hs*/pKJ427 construction was confirmed by PCR as well as *NdeI/XbaI* double digestion and then, the plasmid was transformed into $\Delta htrVIII$ deletion strain using standard transformation protocol. After two week incubation, colonies were picked up and grown in halobacterial growth medium. Each individual culture was checked by PCR to confirm the presence of the plasmid, and by immunoblot to confirm the expression
25 level of HemAT-Hs.

Example 13 – Expression of *hemAT-Bs*

- 30 As with *hemAT-Hs*, three expression systems have been developed. First, *hemAT-Bs* is expressed in the pMAL expression system. In order to express *hemAT-Bs* encoding protein HemAT-Bs in *E. coli*, expression primers were needed to amplify the *hemAT-Bs* gene. Not only the gene, but the ribosomal binding region upstream of the start codon, is required for the expression of HemAT-Bs in *E. coli*.

Table 5: Primers for PCR of HemAT-*Bs*.

Name of Primer	Sequence
<i>hemAT-Bs</i> <i>Bam</i> HI top	ATATGGATCCAAGGGGATCATTGTAATGTTATTAAAAAG SEQ. ID. No. 74
<i>hemAT-Bs</i> <i>Pst</i> I bot	ATTACTGCAGCAACTGATTTTTAATTAAAGTTACATAATGAACGC SEQ. ID. No. 75

5 *Bam*HI and *Pst*I were selected for the cloning of *hemAT-Bs* into expression vector and *E. coli-Bacillus subtilis* shuttle vector. *hemAT-BsBam*HI top/*Pst*I bot primers were used to amplify the *hemAT-Bs* gene from *Bacillus subtilis* genomic DNA by PCR with *pfi* DNA polymerase. After PCR, the amplicon was immediately cloned into TOPO vector using invitrogen TOPO Blunt Cloning kit and transformed into TOP 10 *E. coli* competent cells. Colonies were checked for the right insert.

10 *Bam*HI and *Pst*I were also used for the cloning of the HemAT-*Bs* into pMAL cII vector as well as the shuttle vector pEB 112. *hemAT-Bs/pMAL* construction was transformed to *E. coli* pLysS cells for the expression. After IPTG induction, SDS gel showed two bands in comparison to the uninduced sample. The top band is HemAT-*Bs* protein. The spectra is checked and the results showed clearly the heme protein signature peaks while the MBP itself doesn't show any peak at 410 nm and 541nm/580nm.)

15 Second, the *Nde*I top and *Bam*HI bot primers were used for the cloning of *hemAT-Bs* gene into pET vector. The ribosomal binding region is also included in front of the gene. TOPO cloning was performed after PCR reaction and the construction was confirmed by *Nde*I/*Bam*HI digestion as well as PCR. *hemAT-Bs/pET* construction was transformed into *E. coli* pLysS competent cells. IPTG was used for the protein induction. Spectra showed the specific peaks for heme protein.

20 A peptide consisting of the N-terminal 190 or 250 residues was expressed in pMAL vector. A bottom primer at position 190 and 250 amino acid residue were synthesized with a *Pst*I cutting site. *hemAT-Bs Bam*HI top and these top primers were used to amplify the gene encoding 190 and 250 amino acids at N-terminal of *hemAT-Bs*. The PCR products were cloned into TOPO vector and then subcloned into pMAL HemAT-*Bs* 250 vector by using *Bam*HI/*Pst*I. 190 and 250 *hemAT-Bs/pMAL* constructions were confirmed and transformed into pLysS cells for expression. As

expected, other than MBP, a second protein band appears at position 25 and 30 kDa, which are the sizes of N-terminal 190 and 250 residues of HemAT-*Bs* protein. Spectra also showed the signature peaks of hemeprotein.

A shuttle vector is used for the expression of *hemAT-Bs* gene in its native host
5 *B. subtilis*. *hemAT-Bs*/TOPO construction was used as initial plasmid. The *hemAT-Bs*/pEB 112 construction was transformed into Δ *hemAT-Bs* deletion strain. The transformant was used for physiological study of HemAT-*Bs*.

Example 14 - Construction of a C-terminal His-tag HemAT-*Bs*

10

Two round of *pfu* PCR were performed to generate a C-terminal 6 His-tag to HemAT-*Bs*. The top primer and bottom primer with 6 Histidine codon plus stop codon were used for the first round PCR. The PCR product was cloned into TOPO vector and the resultant vector used for the second round PCR. In the case of pET, *NdeI* and *BamHI*
15 sites were created to clone the insert into expression vector. In the case of pMAL, *BamHI* top and *BamHI* bot primer were used. The final constructions (pET/pMAL) were transformed to *E. coli* pLysS cells for induction.

Example 15 - Site-Directed Mutagenesis of HemAT-*Bs*

20

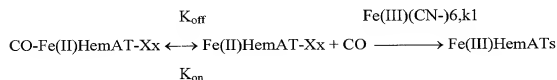
The same strategy is used for generating site-directed mutants for HemAT-*Bs* of *B. subtilis*. The HemAT-*Bs*/TOPO construction with *BamHI* top and *PstI* bottom restriction sites was used as template for PCR-based mutagenesis. HemAT-*Bs* H75A, H86A, H99A, H122A, H123A and H199A are being mutated by PCR-based mutagenesis.
25 The HemAT-*Bs*/pTOPO plasmid was used as initial template for PCR.

The mutants, H75A, H99A, and H123A, have been cloned into pMAL expression vector. H123A spectra showed no significant signature peaks at 540 nm and 580 nm. H123R from pMAL expression culture showed no hemeprotein signature spectra.

Example 16 –Carbon Monoxide Binding in HemAT-*Hs* and HemAT-*Bs*

The rate of CO binding to both HemAT-*Hs* and HemAT-*Bs* was determined by transient absorption spectroscopy using instrumentation described previously (Larsen, et al., *Inorg. Chim. Acta* 234:101-107(1995), which is hereby incorporated by reference).

The rates of CO dissociation for HemAT-*Hs* and HemAT-*Bs* were determined using the ferricyanide method (Gilles-Gonzalez, et al., *Biochemistry* 33:8067-8073 (1994), which is hereby incorporated by reference). Changes in absorbance as a function of time at 418 nm (Soret maximum for the CO bound derivative of each protein) were monitored after the addition of potassium ferricyanide (final concentration of 1.5 mM) to solutions of the co-bound protein. The resulting traces were then fit to single exponential decays to obtain k_{off} assuming the following reaction:



where k_{off}/k_{on} are the dissociation/association rate constants and k_1 is the rate of HemATs oxidation. This procedure relies on k_1 being much larger than k_{off} . In the case of the HemATs proteins this was confirmed by measuring the rate of heme oxidation of the five-coordinate deoxy form the protein.

The optical absorption spectrum of deoxy and CO-bound derivatives of HemAT-*Hs* and HemAT-*Bs* are shown in Figure 4. The absorption spectra of the deoxy forms of both proteins are indicative of five-coordinate high-spin heme with Soret maxima at 425 nm and a broad visible band centered at 555 nm. In the presence of CO the absorption spectrum resembles a six-coordinate low-spin heme with a Soret maximum at ~418 nm (HemAT-*Hs/Bs*) and visible bands at 535 nm and 573 nm.

Figure 6 displays typical transient absorption data subsequent to CO photolysis obtained at 430 nm at 25°C and 1 atm CO for both HemAT-*Hs* (solid line) and HemAT-*Bs* (dotted line). The data can be fit to a single exponential decay indicating a pseudo-first order reaction with CO. The resulting rate constant for CO recombination are found to be $30 \pm 3 \text{ s}^{-1}$ and $132 \pm 3 \text{ s}^{-1}$ for HemAT-*Hs* and HemAT-*Bs*, respectively. Figure 7 shows

the corresponding transient difference spectrum (25 μ s subsequent to photolysis) overlaid with the equilibrium difference spectrum (deoxy minus CO-bound) for HemAT-*Hs* (top panel) and HemAT-*Bs* (bottom panel). The red-shift in the transient difference spectra relative to the equilibrium difference spectra suggest that CO photolysis produces a non-equilibrium five-coordinate complex within 25 μ s subsequent to photolysis.

Figure 8 displays the CO-off rate data for HemAT-*Hs*, HemAT-*Bs*, and horse heart Mb. The CO off-rates are found to be $0.2 \pm 0.01 \text{ s}^{-1}$, $0.098 \pm 0.002 \text{ s}^{-1}$, and $0.056 \pm 0.001 \text{ s}^{-1}$ for HemAT-*Hs*, HemAT-*Bs*, and horse heart Mb, respectively. Using these values along with the second-order rate constants for CO recombination (scaling the pseudo first order rate constants to CO concentration) the associations constants for CO are found to be $1.5 \times 10^4 \text{ M}^{-1}$, $1.35 \times 10^6 \text{ M}^{-1}$, and $7.38 \times 10^6 \text{ M}^{-1}$ for HemAT-*Hs*, HemAT-*Bs*, and horse heart Mb, respectively. These values along with literature values for CO binding to other heme proteins are provided in Table 6.

Table 6: CO-Affinities of various heme proteins.

Protein	K ($\times 10^{-4} \text{ M}^{-1}$)	K _{on} ($\times 10^{-4} \text{ M}^{-1} \text{ s}^{-1}$)	K _{off} (s^{-1})
HemAT- <i>Hs</i> ^a	15	3	0.2
HemAT- <i>Bs</i> ^a	135	13.2	0.098
HH Mb ^a	738	46.5	0.06
SW Mb ^b	2700	51	0.019
SW Mb H(E7)->L ^b	110,000	2,600	0.024
Human HbA ^c	50,000	600	0.013
BjFixL ^c	10	0.5	0.045
RmFixLT ^c	-	1.2	-
RmFixLH ^c	20	1.7	0.083
HRP (pH 7.0) ^c	350	0.3	0.0001
<i>Aplysia</i> Mb ^c	3,000	50	0.02

^a This work.

^b Springer, et al., Chem. Rev. 94:699-714 (1994), which is hereby incorporated by reference.

^c Gilles-Gonzalez, et al., Biochemistry 33:8067-8073 (1994), which is hereby incorporated by reference.

The absorption spectra of oxy-, deoxy-, and carbon monoxide forms of HemAT-*Hs* and HemAT-*Bs* establish that both proteins have a heme prosthetic group to reversibly bind oxygen. Capillary assays demonstrate that both HemAT-*Hs* and HemAT-*Bs* are

involved in negative aerotaxis in phylogenetically distinct archaeon *H. salinarum* and gram-positive bacterium *B. subtilis*, respectively. Thus, the N-terminal segments of HemAT-*Hs* and HemAT-*Bs* may act as sensory domains by binding diatomic oxygen through the heme prosthetic group in the ferrous (Fe(II)) state. This oxygen binding triggers a conformational change in the sensor domain, which in turn alters the activity of the C-terminal signaling domain. This initiates association of the signaling domain with CheW and CheA proteins to generate signals that change the flagellar rotational bias.

Current evolutionary reconstruction indicates that myoglobin, (α - and β -globins derive from a protein that originally appeared in an ancient vertebrate about 500 million years ago (Hardison, Amer. Scientist, 87:126-137 (1999), which is hereby incorporated by reference). However, comparison of amino acid sequences in globins from *Eukarya* and *Bacteria* suggests they share a very early common ancestor, in spite of the fact that the proteins perform different functions (Hardison, Amer. Scientist, 87:126-137 (1999); Hardison, J. Exp. Biol., 201:1099-1117 (1998), which are hereby incorporated by reference). The conserved residues among all myoglobins are the proximal histidine residue in the F helix (F8) and two phenylalanine residues in the CD region (CD1 packs against the heme and CD4 in a hydrophobic cluster in contact with the heme), the distal histidine residue in the E helix (E7) and a proline residue at the beginning of the C helix (C2, sharp turn between B and C helices) (Bashford et al., J. Mol. Biol., 196:199-216 (1987); Vinogradov et al., Comp. Biochem. Physiol., 106B:1-26 (1993), which are hereby incorporated by reference). Three of these residues (proline in C2, phenylalanine in CD4 and histidine in F8) are conserved and phenylalanine in CD1 is replaced by valine in HemAT-*Hs* and HemAT-*Bs* (marked with asterisks in Figure 1A).

HemAT proteins constitute a new class of sensors that differ significantly from the known heme-containing O₂-sensor FixL (16, 17). FixL is a member of the large family of sensor kinases ubiquitous in bacterial two-component regulatory systems. Its heme-binding domain belongs to the PAS-domain superfamily (18, 19). HemATs contain no PAS domains (Taylor, et al., Ann. Rev. Microbiol., 53:90 (1999); Zhulin et al., Mol. Microbiol., 29:1522 (1998), which are hereby incorporated by reference) and differ from FixL both in spectral features and physiological function (Gilles-Gonzalez, et al., Nature, 350:170 (1991); Lois, et al., J. Bacteriol., 175:1103 (1993), which are hereby incorporated by reference). The absorption bands of HemATs are blue-shifted relative to FixL (415 nm Soret band), indicating that the proteins have distinct heme-pocket

geometries. In addition, both HemATs participate in aerotaxis, whereas FixL regulates transcription. HemATs also differ from the aerotaxis transducer Aer in *E. coli*, which has a FAD-binding PAS domain (Rebbapragada et al., Proc. Natl. Acad. Sci. U.S.A., 94:10541 (1997); Bibikov et al., J. Bacteriol., 179:4075 (1997), which are hereby
5 incorporated by reference).

The amino-terminal domains of HemATs are proposed to act as sensors by binding diatomic oxygen at their heme when it is in the ferrous (Fe [II]) state. Oxygen binding presumably triggers a conformational change in the sensor domain that, in turn, alters the activity of the carboxyl-terminal signaling domain. The carboxyl-terminal
10 domains of HemATs are very similar to the signaling domains of the MCP family of bacterial chemoreceptors, which associate with the cytoplasmic CheW and CheA proteins to mediate chemotaxis.

HemATs offer the possibility of being used as biological sensors to monitor physiologically important gases, such as O₂ or CO, because: 1) they are soluble proteins
15 like myoglobin, which has been widely studied at the molecular level; 2) they possess a signaling domain that resembles those of the molecularly well-characterized bacterial chemotaxis transducers; and 3) direct observation of the aerotactic response permits rapid analysis of various perturbations of the sensing and signaling system. In addition, these two proteins provide information about the evolutionary origins of globins in the
20 *Eucarya*, *Archaea*, and *Bacteria*.

Although preferred embodiments have been depicted and described in detail herein, it will be apparent to those skilled in the relevant art that various modifications, additions, substitutions, and the like can be made without departing from the spirit of the invention and these are therefore considered to be within the scope of the invention as
25 defined in the claims which follow.

What is claimed is:

1. An isolated bacterial heme binding protein wherein said protein reversibly binds oxygen with a low affinity and wherein said heme binding domain of said protein
5 shows at least 20% identity to a myoglobin heme binding domain.

2. The isolated heme-binding protein according to claim 1, wherein the protein comprises a heme binding domain and a signaling domain.

10 3. The isolated heme-binding protein according to claim 1, wherein the protein is isolated from *Archaea*.

4. The isolated heme-binding protein according to claim 3, wherein the protein is isolated from *Halobacterium salinarum*.

15 5. The isolated heme-binding protein according to claim 4, wherein the protein's activity is salt tolerant.

6. The isolated heme-binding protein according to claim 1, wherein the
20 protein has an amino acid sequence of SEQ. ID. No. 2.

7. The isolated heme-binding protein according to claim 1, wherein the protein is isolated from *Bacillus subtilis*.

25 8. The isolated heme-binding protein according to claim 7, wherein the protein has an amino acid sequence of SEQ. ID. No. 4.

7. A fragment of the isolated heme-binding protein according to claim 1, wherein said fragment comprises a heme-binding domain.

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8. The fragment according to claim 4, further comprising a heterologous signal transduction domain.

9. A blood substitute comprising:
a bacterial heme binding protein wherein said protein reversibly binds oxygen
with a low affinity.

5 10. The blood substitute according to claim 9, wherein the protein comprises a
heme binding domain and a signaling domain.

11. The blood substitute according to claim 10, wherein the protein is isolated
from *Archaea*.

10

12. The blood substitute according to claim 11, wherein the protein is isolated
from *Halobacterium salinarum*.

13. The blood substitute according to claim 12, wherein the protein's activity
15 is salt tolerant.

14. The blood substitute according to claim 9, wherein the protein has an
amino acid sequence of SEQ. ID. No. 2.

20 15. The blood substitute according to claim 9, wherein the protein is isolated
from *Bacillus subtilis*.

16. The blood substitute according to claim 15, wherein the protein has an
amino acid sequence of SEQ. ID. No. 4.

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17. The blood substitute according to claim 15, comprising a fragment of the
isolated heme-binding protein having a heme-binding domain.

18. The blood substitute according to claim 17, further comprising a
30 heterologous signal transduction domain.

19. A method of treating a patient suffering from low blood levels comprising:
administering to the patient a blood substitute according to claim 9.

20. The method according to claim 19, further comprising:
5 regulating the oxygen binding of the heme-binding protein by modifying the signaling
domain.

21. A method for controlled storage of oxygen, comprising:
providing a bacterial heme binding protein wherein said protein reversibly binds
10 oxygen with a low affinity; and
contacting said protein with oxygen allowing the protein to bind and store oxygen.

22. The method according to claim 21, further comprising:
triggering the release of oxygen from the protein by activating the signaling
15 domain.

23. The method according to claim 21, wherein the protein comprises a heme
binding domain and a signaling domain.

24. The method according to claim 21, wherein the protein is isolated from
Archaea.

25. The method according to claim 24, wherein the protein is isolated from
Halobacterium salinarum.

26. The method according to claim 25, wherein the protein's activity is salt
tolerant.

27. The method according to claim 26, wherein the protein has an amino acid
30 sequence of SEQ. ID. No. 2.

28. The method according to claim 21, wherein the protein is isolated from
Bacillus subtilis.

29. The method according to claim 28, wherein the protein has an amino acid sequence of SEQ. ID. No. 4.

5 30. The method according to claim 21, wherein the protein is a fragment of an isolated bacterial heme binding protein which reversibly binds oxygen with a low affinity, wherein said fragment comprises a heme-binding domain.

31. The method according to claim 30, wherein the fragment further
10 comprising a heterologous signal transduction domain.

32. A method of sensing gaseous ligands comprising:
providing a heme binding bacterial protein wherein said protein reversibly binds
oxygen with a low affinity;
15 exposing said protein to a sample to be tested; and
measuring a change in the conformation of the protein.

33. The method according to claim 32, wherein said measuring is carried out
optically.

20 34. The method according to claim 32, wherein said measuring is carried out electronically.

35. The method according to claim 32, wherein the gaseous ligand is selected
25 from the group consisting of O₂, NO, CO, and CN.

36. The method according to claim 32, wherein the gaseous ligand is O₂.

37. The method according to claim 32, wherein the protein comprises a heme
30 binding domain and a signaling domain.

38. The method according to claim 32, wherein the protein is isolated from
Archaea.

39. The method according to claim 38, wherein the protein is isolated from *Halobacterium salinarum*.

5 40. The method according to claim 39, wherein the protein's activity is salt tolerant.

41. The method according to claim 40, wherein the protein has an amino acid sequence of SEQ. ID. No. 2.

10 42. The method according to claim 32, wherein the protein is isolated from *Bacillus subtilis*.

43. The method according to claim 42, wherein the protein has an amino acid
15 sequence of SEQ. ID. No. 4.

44. The method according to claim 32, wherein the protein is a fragment of an isolated bacterial heme binding protein which reversibly binds oxygen with a low affinity, wherein said fragment comprises a heme-binding domain.

20 45. The method according to claim 44, wherein the fragment further comprising a heterologous signal transduction domain.

46. A chimeric protein comprising:
25 a heme-binding domain of an isolated heme binding bacterial protein; and a heterologous signaling domain.

47. The chimeric protein according to claim 46, wherein the heterologous signaling domain is a mutated signaling domain having altered affinity for its ligand.

30 48. The isolated heme-binding protein according to claim 47, wherein the protein comprises a heme binding domain and a signaling domain.

49. The isolated heme-binding protein according to claim 47, wherein the protein is isolated from *Archaea*.

50. The isolated heme-binding protein according to claim 49, wherein the
5 protein is isolated from *Halobacterium salinarum*.

51. The isolated heme-binding protein according to claim 50, wherein the protein's activity is salt tolerant.

10 52. The isolated heme-binding protein according to claim 51, wherein the protein has an amino acid sequence of SEQ. ID. No. 2.

53. The isolated heme-binding protein according to claim 47, wherein the protein is isolated from *Bacillus subtilis*.

15 54. The isolated heme-binding protein according to claim 47, wherein the protein has an amino acid sequence of SEQ. ID. No. 4.

20 55. An isolated nucleic acid molecule wherein the nucleic acid molecule encodes a heme binding bacterial protein wherein said protein reversibly binds oxygen with a low affinity.

56. The isolated nucleic acid molecule according to claim 55, wherein the nucleic acid molecule comprises:
25 a nucleotide sequence as shown in SEQ. ID. No. 1; or
a nucleotide sequence which hybridizes to a nucleic acid molecule having the sequence shown in SEQ. ID. No. 1 under stringent conditions.

57. A vector comprising the nucleic acid molecule according to claim 55.

30

58. A host cell transformed with the vector according to claim 57.

59. The isolated nucleic acid molecule according to claim 55, wherein the nucleic acid molecule comprises:

a nucleotide sequence as shown in SEQ. ID. No. 3; or

5 a nucleotide sequence which hybridizes to a nucleic acid molecule having the sequence shown in SEQ. ID. No. 3 under stringent conditions.

60. A vector comprising the nucleic acid molecule according to claim 59.

61. A host cell transformed with the vector according to claim 60.

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ABSTRACT OF THE INVENTION

The present invention provides an isolated archaeal and bacterial heme binding protein which reversibly binds oxygen with a low affinity. The heme binding protein may be utilized as a blood substitute. The invention also provides a method for controlled storage of oxygen by contacting a bacterial heme binding protein with oxygen allowing the protein to bind and store oxygen. The also provides methods to sense gaseous ligands using the heme binding protein. In other embodiments, the invention provides chimeric proteins having a heme-binding domain of an isolated heme binding archaeal bacterial protein and a heterologous signaling domain.

[illegible]

|----- A helix -----| |----- B helix -----| | C helix |

SWMb **V**-----**S**CEW**Q**VLHV**A**RV**S**AD**V**A-----**C**H**G**OD**I**LE**L**ES**K**SH**E**PT**L**E**K** 42
HemAT-Hs MSNDNDTLVTADWRNGIDCHALADATGIDP--ASTAWRISFGCHDDDTAAALAEOPLEAT 60
HemAT-Bs **V**EP**K**K**R**K**Q**ET**A**FS**D**SN**G**Q**K**N-R**I**Q**T**N**K**H**A**D**V**K**K**O**L**K**V**R**G**D**A**F**V**LE**S**Q**L**OP**L**Q**E**N 61

| D helix | |----- E helix -----| | F helix

SWMb -----**D**RE**K**HL**I**CE**A**SM**K**AS**E**DL**K**K**E**Q**V**TV**-**-**T**AT**C**N**L**K**K**-**K**CH**H**E**A**S-----**L**K**P****A**Q**S** 92
HemAT-Hs **A**D**A**V**T**D**-**-**V**D**L**ES**V**F**E**Q**D**I**F**AN**S**T**K**V**E**Q**L**K**E**T**A**Y**L**Q**E**CG**E**Y**T**E**Z**A**A**Q**A**R**I**S**K**T 122
HemAT-Bs **H**V**H**I**D**A**-**-**V**R**N**AD**E**SS**I**MD**I**T**-**ND**S**SV**D**RA**K**O**L**K**E**H**I**Q**E**MF**A**G**V**D**E**AT**E**K**S**R**I**A**S** 122

* | |----- G helix -----| |----- H helix -----|

SWMb **A**SK**K**K**K**L**I**K**V**T**E**SE**A**TE**H**VL**H**SR**H**PG**D**SC**A**Q**C**AN**N**AL**E**LF**R**K**O**TA**A**-**K**Y**K**E**L**Q**V**Q**G** 153
HemAT-Hs **H**D**V**GL**E**Q**V**AL**G**A**T**AT**G**CL**D**AD**D**V**Q**RG**E**SA**A**Q**D**SL**V**ARE**L**EM**K**ET**T**SD**Q**Q**H** 184
HemAT-Bs **H**TE**L**GL**E**K**V**Y**H**GA**Q**Q**S**TA**S**MT**Q**Y-----**S**SL**S**Q**Q**Q**L**KA**R**AT**T**K**T**NG**Q**Q**L** 175

SWMb SEQ. ID. No. 76
HemAT-Hs SEQ. ID. No. 77
HemAT-Bs SEQ. ID. No. 78

a

* * * K1 * * *

Tsr **L**MR**V**Q**D**VR**N**GA**N**AL**Y**SC**A**SE**L**IT**G**NN**D**VS**R**TP**Q**AS**L**E**E**TA**S**ME**O**L**T**AT**V**K**N**AE**N**E**R** 324
HemAT-Hs **L**EA**R**SC**D**VA**E**RT**D**T**R**A**R**AT**D**Q**D**RM**A**D**V**SR**E**TS**S**VS**A**SV**E**VA**S**T**A**DD**V**RR**R**SE**D**AE**A**U**A**Q 283
HemAT-Bs **L**H**K**Q**I**Q**E**TS**G**SH**A**N**L**ES**E**TS**R**SV**O**EL**V**DK**SE**Q**S**Q**A**SK**A**Q**E**TV**S**ST**V**EE**K**ST**G**CK**R**EE**V**Q**Q** 259

HCD

Tsr **Q**ASH**L**AL**S**AGE**TA**ORG**C**K**V**DN**V**Q**T**MR**D**TS**S**SK**I**AD**IS**VID**G**IA**F**Q**T**N**L**AL**N**A**V**EA 386
HemAT-Hs **Q**SE**A**AD**D**AL**A**TT**Q**ID**E**AT**D**Q**Y**T**A**G**VE**Q**SR**RA**D**VE**S**TV**E**VID**D**IA**R**Q**T**N**L**AL**N**AS**T**EA 345
HemAT-Bs **K**-----**Q**N**K**ID**TS**H**V**Q**IS**E**K**W**R**Q**D**HA**Q**LE**K**IP**GV**TV**G**IA**R**Q**T**N**L**SL**N**AS**IE**S 311

HCD

Tsr **A**RAGE**Q**RC**G**PA**V**VA**Q**EV**N**LA**Q**SA**A**AN**PI**HS**L**TS**SV**CH**V**Y**VS**SL**VE**Q**CT**MA**EV**Q**A** 448
HemAT-Hs **A**RAGE**Q**RC**G**PA**V**VA**Q**EV**A**LA**D**ES**R**Q**SP**FA**VE**LV**Y**2**HQ**AN**TS**Q**VD**Q**D**Q**W**RA**K**Q**SE**Q**V**R 407
HemAT-Bs **A**RAGE**Q**RC**G**PA**V**VA**Q**EV**E**K**L**SE**ET**KK**V**TS**VS**EL**V**-----**N**Q**N**Q**Q**Q**TV**VS**K**IA**K**Q**Q**N**ST** 366

* * K1 * * *

Tsr **V**SE**V**Q**D**TS**Q**IAS**Q**SE**D**ES**R**Q**ID**Q**V**GL**V**AE**MD**RV**Q**Q**AL**VE**S**SA**A**AA**AA**AE**Q**Q**N**SL**SE** 510
HemAT-Hs **V**SE**A**Q**Q**SL**Q**IT**DA**VED**A**SC**Q**EW**ST**AT**D**Q**AV**SE**EV**AE**MD**Q**Q**DR**AGE**TA**AA**AT**D**IA**D** 469
HemAT-Bs **V**SE**S**Q**Q**Q**Q**Q**Q**IN**AN**Q**VD**EL**V**Y**S**MS**IS**KE**Q**SE**K**ID**Q**Q**Q**AL**LG**Q**Q**Q**Q**SV**RA**VS**HA**AS**VD**SL**VT** 428

Tsr **A**V**A**VF**R**IQ**Q**Q**Q**RE**S**AV**V**RE**TV**Q**AMP** SEQ. ID. No. 79 536
HemAT-Hs **A**T-----**Q**Q**Q**V**RE**VE**V**RE**TV**Q**K**LS SEQ. ID. No. 80 489
HemAT-Bs **AT**-----**Q**Q SEQ. ID. No. 81 432

b

FIGURE 1

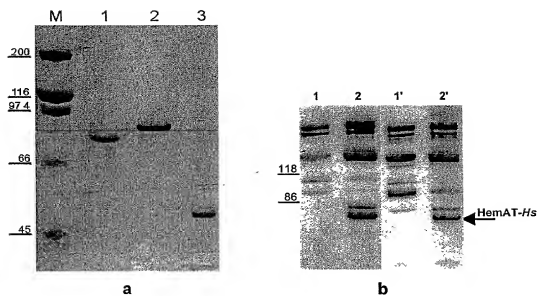


FIGURE 2

Myoglobin-like Protein (MbLP)

QQDVLVVLKXHPLIQEKIXFDFFKR

XXXXXXXXXXXXXXXXXXXXXXXXXXXX

AQRXRLAQIHAXKGKIPDWYL

M1-box

SEQ. ID. No. 82

M2-box

SEQ. ID. No. 83

TEMPLATE

IIEXTVFVLXEHGXII

QQDVLVVLKXNPEIQEKFFFFKR

XXXXXXXXXXXXXXXXXXXXXXXXXXXX

AQRXRLAQIHAXKGKIPDWYL

H-box

SEQ. ID. No. 84

M1-box

SEQ. ID. No. 85

M2-box

SEQ. ID. No. 86

Figure 1 The two sequences used in the analyses. M1 and M2 are the site of myoglobin recognition. M2 is the site of HemAT recognition. The H-box is the site primarily of microbial hemoglobin recognition.

FIGURE 3

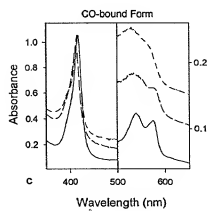
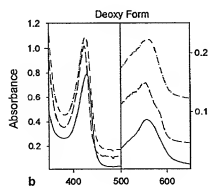
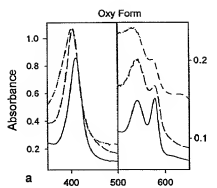


FIGURE 4

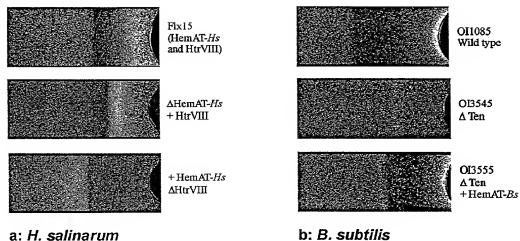


FIGURE 5

00001-8205424

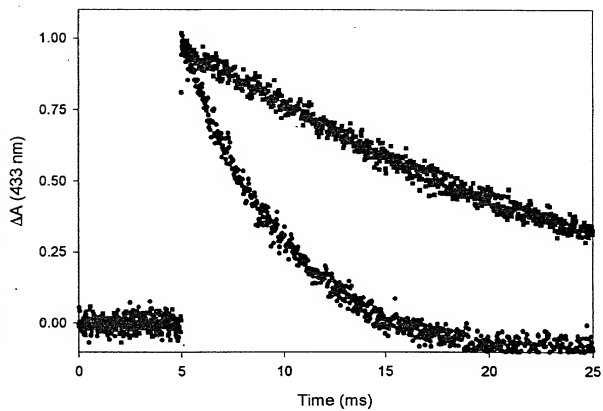


FIGURE 6

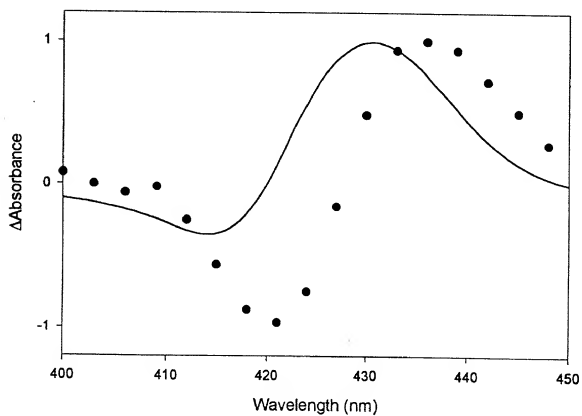
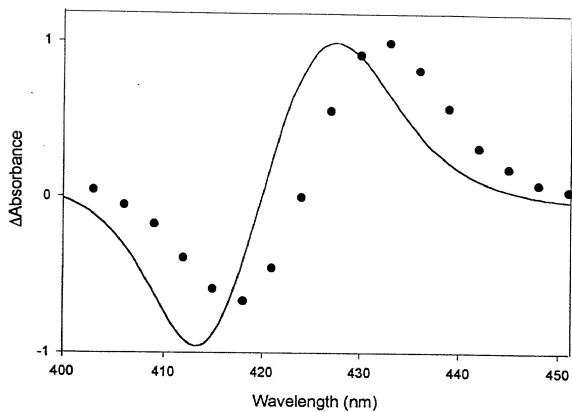


FIGURE 7

COMBINED DECLARATION FOR PATENT
APPLICATION AND POWER OF ATTORNEY
(Includes Reference to PCT International Applications)

ATTORNEY'S DOCKET NUMBER
201040/1020

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

HEME PROTEINS HEMAT-*HS* AND HEMAT-*BS* AND THEIR USE IN MEDICINE AND MICROSENSORS

the specification of which (check only one item below):

☒ is attached hereto.

☐ was filed as U.S. Patent Application Serial No. _____ on _____ and was amended on _____ (if applicable).

☐ was filed as PCT International Application Number _____ on _____ and was amended under PCT Article 19 on _____ (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specifications, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the patentability of this application in accordance with Title 37, Code of Federal Regulations, § 1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, § 119 of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States listed below and have also identified below any foreign application(s) for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) of which priority is claimed:

FOR FOREIGN/PCT APPLICATION(S) AND ANY PRIORITY CLAIMS UNDER 35 U.S.C. 119:

COUNTRY (IF PCT, indicate "PCT")	APPLICATION NUMBER	DATE OF FILING (day, month, year)	PRIORITY CLAIMED UNDER 35 USC 119
			<input type="checkbox"/> YES <input type="checkbox"/> NO
			<input type="checkbox"/> YES <input type="checkbox"/> NO
			<input type="checkbox"/> YES <input type="checkbox"/> NO
			<input type="checkbox"/> YES <input type="checkbox"/> NO
			<input type="checkbox"/> YES <input type="checkbox"/> NO
			<input type="checkbox"/> YES <input type="checkbox"/> NO
			<input type="checkbox"/> YES <input type="checkbox"/> NO
			<input type="checkbox"/> YES <input type="checkbox"/> NO

**COMBINED DECLARATION FOR PATENT
APPLICATION AND POWER OF ATTORNEY (Continued)
(Includes Reference to PCT International Applications)**

ATTORNEY'S DOCKET NUMBER
201040/1020

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s) or PCT international application(s) designating the United States of America that is/are listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in that/those prior application(s) in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, § 1.56(a) which occurred between the filing date of the prior application(s) and the national or PCT International filing date of this application:

PRIOR U.S. APPLICATIONS OR PCT INTERNATIONAL APPLICATIONS DESIGNATING THE U.S. FOR BENEFIT UNDER 35 U.S.C. 120:

U.S. APPLICATIONS		STATUS (Check One)		
U.S. APPLICATION NUMBER	U.S. FILING DATE	PATENTED	PENDING	ABANDONED
PCT APPLICATIONS DESIGNATING THE U.S.				
PCT APPLICATION NO.	PCT FILING DATE	U.S. SERIAL NUMBERS ASSIGNED (if any)		

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith. **Michael L. Goldman, Registration No. 30,727; Gunnar G. Leinberg, Registration No. 35,584; Dennis M. Connolly, Registration No. 40,964; Edwin V. Merkel, Registration No. 40,087; Georgia Caton, Registration No. 44,597**

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1 2 3 4 5 6 7 8 9 10 11 12	FULL NAME OF INVENTOR	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME
	RESIDENCE & CITIZENSHIP	CITY	STATE/FOREIGN COUNTRY	COUNTRY OF CITIZENSHIP
	POST OFFICE ADDRESS	P.O. ADDRESS	CITY	STATE & ZIP CODE/COUNTRY
2 0 2	FULL NAME OF INVENTOR	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME
	RESIDENCE & CITIZENSHIP	CITY	STATE/FOREIGN COUNTRY	COUNTRY OF CITIZENSHIP
	POST OFFICE ADDRESS	P.O. ADDRESS	CITY	STATE & ZIP CODE/COUNTRY
2 0 3	FULL NAME OF INVENTOR	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME
	RESIDENCE & CITIZENSHIP	CITY	STATE/FOREIGN COUNTRY	COUNTRY OF CITIZENSHIP
	POST OFFICE ADDRESS	P.O. ADDRESS	CITY	STATE & ZIP CODE/COUNTRY

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

SIGNATURE OF INVENTOR 201 UNSIGNED	SIGNATURE OF INVENTOR 202 UNSIGNED	SIGNATURE OF INVENTOR 203
DATE	DATE	DATE